

Chalk rocky grasslands in the arid Eastern European Plain

Kalkfelsrasen in der trockenen osteuropäischen Tiefebene

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Abstract

Chalk rocky grasslands of the arid Eastern European Plain are endangered and highly important for biodiversity protection. This study aimed to revise previous syntaxonomic concepts, compare chalk rocky grasslands with other calcareous rocky grasslands and provide a unified classification of chalk rocky grasslands. We extracted relevés from the European Vegetation Archive and added relevés from protologues and papers describing chalk rocky grasslands within the study area. We pre-selected an initial dataset of calcareous rocky grasslands, which we narrowed down to the order *Stipo pulcherrimae-Festucetalia pallentis*. These were further classified using a modified TWINSpan algorithm. We classified them as *Anabasio cretaceae-Agropyron desertori*, *Sileno fruticosae-Nanophytion erinacei*, *Anthemido trozkianae-Artemision salsoloides*, *Sileno supinae-Artemision hololeuca*, *Euphorbio cretophilae-Thymion cretacei* and transitional communities of the *Centaureo carbonatae-Koelerion talievii* alliances within two orders with validation of some syntaxa. The communities are clearly differentiated from each other along gradients of climate and edaphic properties. Differences to other European limestone communities are given. We provide the first unified European classification of continental chalk rocky grasslands, including Ukrainian, Russian and Kazakh communities.

Keywords: chalk rocky grasslands, vegetation, Eastern European Plain, Ukraine, Kazakhstan

Erweiterte deutsche Zusammenfassung am Ende des Artikels

1. Introduction

Chalk rocky grasslands dominated by specific endemic chamaephytes and hemi-cryptophytes with star-shaped and cobwebby pubescence occur in eastern Europe's chalk outcrops of river valleys. They occupy skeletal soils (rendzic, calcareous and calcic leptosols) and strongly depend on the high carbonate content of the chalk bedrock.

The vegetation of chalk outcrops has been studied since the late 19th century and has been the subject of numerous discussions, particularly regarding its classification, structure, and origin. Litvinov (1891, 1902) initially characterized chalk rocky grasslands. Later, Kozo-Polianskyi (1931) distinguished two components within it: the “hyssop flora” (*Hys-sopus cretaceus*, *Artemisia hololeuca*, *A. salsoloides*), which has a relict character and is genetically related to the southern flora of the Central and Middle Asia, and the flora of

“lower alpine” habitats, associated with northern Periglacial steppes (e.g. *Carex humilis*, *Androsace kozo-poljanskii*). Chalk rocky grasslands, unlike steppes dominated by hemi-cryptophytes with the high presence of graminoids, are characterized by a sparse cover (up to 60%) consisting mainly of chamaephytes, forming on skeletal soils without continuous dead litter. Taxonomically, many genera of *Thymus*, *Helianthemum*, *Artemisia*, *Hyssopus*, *Scrophularia*, *Pimpinella*, and *Asperula*) are subdivided into sections, subsections, series, and numerous “minor” vicarious species endemic to specific regions, indicating the uniqueness of this flora and progressive evolution of speciation (Lavrenko 1961). This distinctiveness from steppe vegetation was reflected in dominant classifications, with this vegetation being treated as thyme communities (Lavrenko 1961) or tomilliaries (Didukh 1981, 1989), analogues of Balkan phrygana.

A classification of the chalk rocky steppe vegetation of the Central Russian Upland (Ukraine and part of Russia) was first provided by Didukh (1989), who described a new order of *Thymo cretacei-Hyssopetalia cretacei* and lower syntaxa within the *Festuco-Brometea* class. Romashchenko et al. (1996) described chalk rocky vegetation within a new class of *Helianthemo-Thymetea* that was not floristically similar to pure steppe vegetation, in which the *Thymo cretacei-Hyssopetalia cretacei* order, *Artemisio hololeuca-Hyssopion cretacei*, *Euphorbio cretophilae-Thymion cretacei* and *Centaureo carbonatae-Koelerion talievii* alliances were considered. Further studies of chalk rocky vegetation in Ukraine and Russia have been analyzed in detail by Didukh et al. (2018); however, the similarity of chalk and limestone rocky grasslands led to misinterpretation in Moldova (Rushchuk et al. 2005, Pinzaru 2006).

Chalk rocky vegetation has also been described within *Anabasieta cretaceae* (Golovanov et al. 2021) east of the Volga River in the steppe region that transforms into a semi-desert and desert region in the south, which has specific desert-steppe, North Turanian steppe species (Darbaieva 2003, Riabinina & Lukianova 2005, Saksonov et al. 2019, Golovanov et al. 2021).

Generally, Mucina et al. (2016) consider these communities within the zonal steppe vegetation of the *Festuco-Brometea* class, which spreads along the temperate belt and varies from Mediterranean wet to Asian aridic conditions.

However, the absence of *Festuco-Brometea* diagnostic species causes a heated discussion about their modern syntaxonomy. The *Centaureo carbonatae-Koelerion talievii* alliance has been considered within the *Festucetalia valesiacae* or *Stipo pulcherrimae-Festucetalia pallentis* orders due to the floristic and ecological similarities of these communities with rocky grasslands (Mucina et al. 2016, Didukh et al. 2021).

Chalk rocky grasslands are currently endangered and highly important for biodiversity protection. The main threats are afforestation, terracing of the slopes and the mining of chalk and marl (Kuzemko 2022). Since the Russian aggression in 2014, new threats have arisen that influence the endangered chalk rocky grasslands: direct vanishing of the vegetation cover due to intensive military activities, abandoned agricultural lands near the slopes of chalk outcrops and proliferation of invasive species. Nevertheless, in the Bern Convention, they are of conservation importance in Europe as E1.13 Continental dry, rocky steppe grasslands and dwarf scrub on chalk outcrops (Schaminée et al. 2016).

The main goal of the current study was to revise the classification of chalk rocky vegetation within the Eastern European Plain based on a large international dataset, and to complete knowledge of its syntaxonomy, ecology and distribution. To achieve this goal, we aimed to (1) revise the syntaxonomy of chalk rocky vegetation within the Eastern European

Plain and compare it with other types of calcareous rocky grasslands; (2) analyse the distribution of chalk rocky grasslands within the study area; (3) characterise the main environmental factors that cause the differentiation of chalk rocky grasslands.

2. Study area

We considered rocky grasslands on chalk outcrops within the aridic zone of the Eastern European Plain (Fig. 1). Specifically, we focused on two distinct regions: (1) the Central Russian Upland, and (2) the Sub-Ural Upland across a climatic gradient that significantly impacts the distribution and composition of vegetation types within the study area in which chalk outcrops occur.

The Central Russian Upland occupies eastern Ukraine and southwestern Russian Federation, extending from northwest to southeast, from the right bank of the Oka to the Donets River, with an elevation of 230–250 m a.s.l. (Anon 2007). The Central Russian Upland is built of Precambrian deposits of the crystalline Voronezh Massif covered by layers of sedimentary deposits of the Devonian, Jurassic, Cretaceous and Paleogene periods (Anon 2007). The Central Russian Upland has a sub-continental (subaridic) climate, characterised by large temperature amplitudes between winter and summer. It generally has 450–500 mm annual precipitation, with an average temperature of around -4 – 6 °C in January and $+21$ – 24 °C in July, with essential temperature amplitudes (27 – 30 °C), Gorchinskyi continentality coefficient ($I_g = 42$ – 46), de Martonne coefficient of aridity-humidity ($I_{dm} = 28,5$ – 25), with a wetter climate in the western parts of the region and a drier climate in the east (Karger et al. 2017, Didukh 2023).

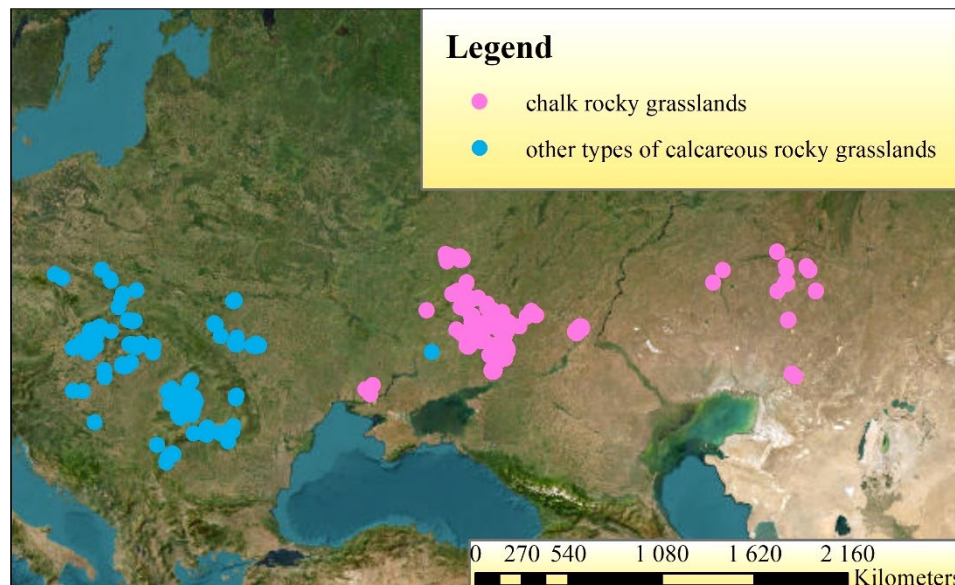


Fig. 1. Map of the chalk rocky grasslands compared with other types of calcareous rocky grasslands distribution.

Abb. 1. Karte der untersuchten Kalkfelsrasen im Vergleich zu anderen Typen von Kalkfelsrasen.

The Sub-Ural Upland is a geological and geographical region occupied by the Aktobe region in Kazakhstan, with a northwestern part in the West Kazakhstan Region and a small northern part in the Orenburg region (Russian Federation) with an elevation of 100–450 m a.s.l. (Anon. 2010). The Sub-Ural Upland predominantly comprises ancient crystalline rocks, including granite and gneiss, formed during the Precambrian period. It also contains upper layers of sedimentary bedrock, such as limestone, chalk and sandstone, deposited during later geological periods. The climate in the Sub-Ural Upland region varies depending on the location and elevation. Generally, it experiences a subcontinental climate with moderate annual precipitation, which typically ranges from 250 to 400 mm, with slightly higher amounts in the western parts of the upland, with an average temperature of around -9–13 °C in January and +22–25 °C in July, with essential temperature amplitudes (30–36 °C), Gorchinskyi continentality coefficient ($I_g = 46\text{--}60$), de Martonne coefficient of aridity-humidity ($I_{dm} = 25\text{--}20$) (Karger et al. 2017, Didukh 2023).

3. Materials and methods

3.1 Vegetation data for broad-scale comparison

To revise the high-level syntaxonomical units of chalk rocky vegetation, we digitised data from the literature or extracted it from EVA and used it for comparison with data from the study area and adjacent territories.

We selected the data from the literature with original diagnoses of the *Anabasietaea cretaceae* class Golovanov et al. 2021 (146 relevés), the “*Helianthemo-Thymetea* class Romashchenko et al. 1996” (225 relevés), the “*Thymo cretacei-Hyssopetalia cretacei* order Didukh 1989” (90 relevés) and subordinate syntaxa (181 relevés) (Sereda 2008, 2009, Demina 2011, Poluyanov & Averinova 2012, Averinova 2014, Koroliuk et al. 2021).

In the EVA database (Chytrý et al. 2016), we also selected all relevés of calcareous rocky grasslands from Poland (87 relevés), Slovakia (250 relevés), Hungary (751 relevés), Romania (357 relevés), Ukraine and Moldova (580 relevés) assigned to the *Stipo pulcherrimae-Festucetalia pallentis* order Pop 1968 or subordinate syntaxa, by the initial authors or custodians of the databases. Additionally, we created a database of the Central Russian Upland (504 relevés) and added it to the general dataset.

We unified the taxonomy of vascular plants according to Euro+Med Plantbase (Euro+Med 2006+) and, in addition, Cherepanov’s checklist (1995), bryophytes according to Hodgetts et al. (2020), and lichens according to the Index Fungorum (2021). We combined them into aggregates (Supplement E1), removed bryophytes and lichens because they were not recorded in some relevés, and returned them to the final synoptic table (Supplement E2).

3.2 Expert system processing

From the initial dataset of 3171 relevés, we selected relevés assigned as EUNIS habitats R1C, R15, R16 and R based on the EUNIS-ESy expert system (Chytrý et al. 2020). Afterwards, we checked these relevés by the expert system of Willner et al. (2017) and extracted data assigned to the *Stipo pulcherrimae-Festucetalia pallentis* order. Other relevés, such as other EUNIS habitat codes and *Festuco-Brometea* orders, were removed from the analysis. After expert processing, we got a dataset of 1643 relevés of the calcareous vegetation.

3.3 TWINSpan classification

In our analysis, we applied a modified TWINSpan algorithm (Hill 1979, Roleček et al. 2009), utilising three pseudospecies cut levels of 0%, 5% and 25% cover. The minimum group size for the division was set at five relevés, and Whittaker’s beta coefficient served as a metric for internal cluster

heterogeneity. We assessed the optimal number of clusters using OptimClass 1 (Tichý et al. 2010) on the first stage of the analysis.

On reviewing the classification results, we employed non-modified TWINSpan (Hill 1979) to classify three clusters separately, in view of their ecological and floristic heterogeneity. The same pseudospecies cut levels (0%, 5% and 25% cover) were used, with a minimum group size for division set at ten relevés and two division levels.

Diagnostic species for each cluster were identified based on the phi-coefficient of association and applied to virtually standardised cluster sizes (Chytrý et al. 2002, Tichý & Chytrý 2006). A fidelity threshold of $\phi = 0.30$ was used, together with Fisher's exact test ($p < 0.05$).

3.4 Ecological indicators and distribution of the communities

We used EIVE 1.0 scales (Dengler et al. 2023) to find environmental differences between main orders of calcareous rocky grasslands in the broad-scale comparison context.

In addition, we used Didukh's scales to explore environmental differences in chalk rocky grasslands within the study area precisely. The phytosociological scales of Didukh (2011) reflect amplitude indices of the species characterised by the following dimensions: ecological indicator of soil humidity (Hd – 23 grades), ecological indicator of variability of damping (fH – 11 grades), ecological indicator of soil acidity (Rc – 15 grades), ecological indicator of total salt regime (Sl – 19 grades), ecological indicator of carbonate content (Ca – 13 grades), ecological indicator of nitrogen content (Nt – 11 grades), ecological indicator of soil aeration (Ae – 15 grades), ecological indicator of thermoregime (Tm – 17 grades), ecological indicator of humidity of microclimate (Om – 23 grades), ecological indicator of continentality (Kn – 17 grades), ecological indicator of cryoregime (Cr – 15 grades) and ecological indicator of light in the community (Lc – 9 grades).

For each relevé, we calculated non-weighted means of species' values for the environmental parameters and fitted them to the Detrended Correspondence Analysis (DCA) ordination plot using *envfit* function of the *vegan* R package. DCA was also calculated using *vegan*, with default parameters. We transformed species abundance data from the seven-grade Braun-Blanquet cover-abundance scale to percentage covers as follows: r = <1%, + = <2%, 1 = <3%, 2 = 13%, 3 = 38%, 4 = 68%, 5 = 88%. The obtained values were then log-transformed as $\log(x + 1)$ to diminish distribution skewness (Borcard et al. 2018, Tichý et al. 2020).

Depending on anatomical and morphological features, physiological and biological properties, growth processes, plant development, preservation and transmission of genetic information, three ecological strategies were distinguished: CRS, which reflects assimilation, accumulation and transfer of resources, and energy. To define the strategies of species on the phytosociological level, we constructed a Grime's triangle using Didukh's methodology (Supplement E3).

Based on the predominance of the species strategy types in the communities, we considered three types of habitats: SPE (successive ecosystems), C – concurrent strategy PSE (persistent stable ecosystems), S – stress-tolerance strategy and FLE (fluctuation labile ecosystems), R – ruderal strategy (Didukh 2023). This approach makes it possible to evaluate the ecological strategy from the standpoint of functioning, specifics of development, dynamics and stability of ecosystems, in particular conditions of existence, which assimilate, accumulate and transfer resources and energy, and preserve and ensure the spread of genetic material (Didukh 2023).

In addition, we classified all the vascular plants into one of the Raunkiaer plant life forms: therophytes, geophytes, hemicryptophytes, chamaephytes and phanerophytes (Raunkiaer 1934). The data are available in Supplements E2.

3.5 Applications and Software

The results were visualized using *vegan* version 2.6-4 (Oksanen 2022) and *ggplot2* (Wickham 2016) packages in R (R Core Team 2021). The TWINSpan classification was done with JUICE version 7.1 (Tichý 2002). Maps were created using ArcGIS 10.8.2. We used the following sources as the basemaps: National Geographic World Map, World Imagery, Biomes and Ecoregions 2017 (<https://services.arcgis.com>).

4. Results and Discussion

4.1 Cluster results of the initial vegetation dataset

We obtained 8 clusters after the suggestion of Optimclass 1 (Tichý et al. 2010) using the modified TWINSpan algorithm (Roleček et al. 2009) in the first stage of the analysis. In the second stage of the analysis, we classified the first, second and last clusters separately due to the floristic heterogeneity of their constructions using the original TWINSpan algorithm (Hill 1979) and obtained thirteen clusters (Fig. 2). The first six clusters (also Table 1) that contained chalk rocky grasslands (482 relevés) are described in detail below. The 7–13 clusters (1161 relevés) corresponding to the order *Stipo pulcherrimae-Festucetalia pallentis* that is not in the scope of our paper and used for comparison of life forms and ecological strategies are shown in supplementary materials (Supplement E4).

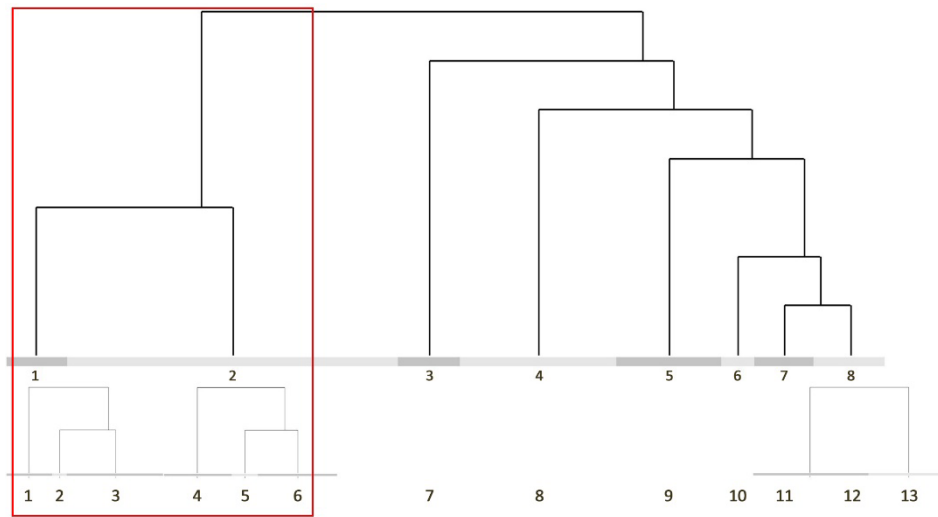


Fig. 2. Dendrogram of chalk rocky grasslands and other calcareous rocky grasslands based on the TWINSpan algorithm. The upper dendrogram represents the first stage (eight clusters were obtained), and the lower diagrams represent the second stage (thirteen clusters were obtained). The chalk rocky grasslands are highlighted with the red frame. Non-highlighted clusters represent the other types of calcareous rocky grasslands.

Abb. 2. Dendrogramm der untersuchten und anderen Kalkfelsrasen basierend auf dem TWINSpan-Algorithmus. Das obere Dendrogramm stellt die erste Stufe dar (es wurden acht Cluster erhalten), und die unteren Diagramme stellen die zweite Stufe dar (es wurden dreizehn Cluster erhalten). Die untersuchten Kalkfelsrasen sind mit einem roten Rahmen hervorgehoben, während die nicht hervorgehobenen Cluster die anderen Typen von Kalkfelsrasen darstellen.

Table 1. The shortened synoptic table shows the percentage constancy of species in six clusters that represent chalk rocky grasslands. Species are ranked by decreasing fidelity measured with the *phi*-coefficient, which is not shown. Species with a *phi*-value greater than 0.30 are considered diagnostic and shaded in light grey; species with a *phi*-value greater than 0.50 are considered highly diagnostic and shaded in dark grey. Species highly diagnostic or diagnostic in one cluster are accepted as diagnostic for alliances; species highly diagnostic or diagnostic in more than one cluster are accepted as diagnostic for high-rank syntaxa. Species with a constancy of 15% or less in individual columns are not shown in the table unless they are diagnostic.

Tabelle 1. Die gekürzte synoptische Tabelle zeigt die prozentuale Stetigkeit der Arten in sechs Clustern von Kalkfelsrasen. Die Arten sind nach abnehmender Treue geordnet, gemessen mit dem nicht angegebenen *phi*-Koeffizienten. Arten mit einem *phi*-Wert größer als 0,30 gelten als diagnostisch und sind hellgrau schattiert; Arten mit einem *phi*-Wert größer als 0,50 gelten als hochdiagnostisch und sind dunkelgrau schattiert. Arten, die in einem Cluster hochdiagnostisch oder diagnostisch sind, werden als diagnostisch für Verbände akzeptiert; Arten, die in mehr als einem Cluster hochdiagnostisch oder diagnostisch sind, werden als diagnostisch für hochrangige Syntaxa akzeptiert. Arten mit einer Stetigkeit von 15 % oder weniger in einzelnen Spalten werden in der Tabelle nicht angezeigt, es sei denn, sie sind diagnostisch.

ID clusters	1	2	3	4	5	6
Number of relevés	35	11	73	243	92	28
All. <i>Anabasio cretaceae</i>-<i>Agropyron desertori</i>						
<i>Astragalus kustanaicus</i>	63	.	.	.	.	.
<i>Bassia prostrata</i>	89	9	19	5	2	2
<i>Sterigmotemum caspicum</i>	69	9	7	.	.	.
<i>Galatella tatarica</i>	57	.	4	.	.	.
<i>Agropyron cristatum</i> aggr.	94	.	52	1	1	6
<i>Krascheninnikovia ceratoides</i>	63	.	7	5	3	1
<i>Tulipa scythica</i>	49	.	5	.	.	.
<i>Lappula microcarpa</i>	63	.	27	.	.	.
<i>Allium tulipifolium</i>	43	.	7	.	.	.
<i>Tulipa biflora</i>	37	.	1	.	.	.
<i>Tulipa sylvestris</i>	37	.	1	.	.	.
<i>Tragopogon ruber</i>	40	.	5	.	.	.
<i>Poa bulbosa</i>	40	.	4	1	.	5
<i>Taraxacum</i> sect. <i>Macrocornuta</i>	31	.	1	.	.	.
<i>Isatis gymnocarpa</i>	29	.	.	.	.	.
<i>Asparagus inderiensis</i>	34	.	7	.	.	.
<i>Meniocus linifolius</i>	37	.	1	.	.	1
<i>Litwinowia tenuissima</i>	23	.	.	.	.	.
<i>Stipa sareptana</i>	46	18	12	.	.	.
<i>Caragana balchaschensis</i>	23	.	1	.	.	.
<i>Ferula caspica</i>	23	.	3	.	.	.
<i>Palimbia turgaica</i>	23	.	3	.	.	.
<i>Gagea bulbifera</i>	17	.	.	.	.	.
<i>Pseudosedum lievenii</i>	17	.	.	.	.	.
<i>Rochelia disperma</i> subsp. <i>retorta</i>	17	.	.	.	.	.
<i>Limonium meyeri</i>	17	.	.	.	.	.
<i>Allium lineare</i>	17	.	.	.	.	.
<i>Rindera tetraspis</i>	31	18	.	.	.	.
<i>Artemisia semiarida</i>	31	18	3	.	.	.
<i>Eriosynaphe longifolia</i>	14	.	.	.	.	.
<i>Astragalus temirensis</i>	14	.	.	.	.	.
<i>Tulipa suaveolens</i>	14	.	.	.	.	.
<i>Anabasis salsa</i>	23	9	.	.	.	.
<i>Filago arvensis</i>	14	.	.	.	.	.
<i>Klasea erucifolia</i>	14	.	.	.	.	.

ID clusters	1	2	3	4	5	6
Number of relevés	35	11	73	243	92	28
<i>Silene otites</i> aggr.	14	.	.	.	.	1
<i>Linaria incompleta</i>	20	.	7	.	.	.
<i>Artemisia lessingiana</i>	29	.	21	.	.	.
<i>Androsace maxima</i>	17	.	5	.	.	.
<i>Allium inderiense</i>	11	.	.	.	.	.
<i>Silene viscosa</i>	11	.	.	.	.	.
<i>Takhtajaniantha pusilla</i>	11	.	.	.	.	.
<i>Ixiolirion tataricum</i>	11	.	.	.	.	.
<i>Ornithogalum fischerianum</i>	11	.	.	.	.	1
All. <i>Sileno fruticosae</i>-<i>Nanophyton erinacei</i>						
<i>Matthiola tatarica</i>	.	100	.	.	.	.
<i>Gypsophila diffusa</i>	.	100	.	.	.	.
<i>Onosma staminea</i>	.	91	.	.	.	.
<i>Nanophyton erinaceum</i>	11	91	18	.	.	.
<i>Atraphaxis spinosa</i>	.	64	.	.	.	.
<i>Artemisia terrae-albae</i>	3	64	.	.	.	.
<i>Atraphaxis replicata</i>	3	64	.	.	.	.
<i>Galium humifusum</i>	.	64	.	1	.	3
<i>Silene antri-jovis</i>	.	55	4	.	.	.
<i>Lepidium meyeri</i>	.	73	25	4	.	.
<i>Lomelosia isetensis</i>	6	73	25	.	.	.
<i>Anabasis cretacea</i>	46	100	6	.	.	.
<i>Astragalus medius</i>	3	36	1	.	.	.
<i>Astragalus stenoceras</i>	.	27	.	.	.	.
<i>Rhammatophyllum pachyrhizum</i>	.	27	1	.	.	.
<i>Limonium suffruticosum</i>	3	27	.	.	.	.
<i>Ephedra distachya</i>	34	73	47	.	9	3
<i>Convolvulus fruticosus</i>	.	18	.	.	.	.
All. <i>Anthemido trotzkiana</i>-<i>Artemision salsoloides</i>						
<i>Archanthemis trotzkiana</i>	6	.	90	.	.	.
<i>Zygophyllum pinnatum</i>	.	9	67	.	.	.
<i>Atraphaxis decipiens</i>	.	.	48	.	.	.
<i>Matthiola fragrans</i>	3	.	82	45	10	6
<i>Astragalus tenuifolius</i>	23	.	56	.	.	.
<i>Hedysarum tscherkassovae</i>	3	.	36	.	.	.
<i>Jurinea kirghisorum</i>	.	.	25	.	.	.
<i>Gypsophila rupestris</i>	.	.	21	.	.	.
<i>Artemisia salsoloides</i>	9	.	74	33	53	15
<i>Hedysarum razoumovianum</i>	3	.	18	.	.	.
<i>Gypsophila altissima</i>	.	.	12	.	.	.
<i>Galium octonarium</i>	9	.	27	9	1	5
<i>Atraphaxis frutescens</i>	.	.	14	2	.	.
All. <i>Sileno supinae</i>-<i>Artemision hololeuca</i>						
<i>Hyssopus officinalis</i>	.	.	.	81	8	10
<i>Scrophularia cretacea</i>	.	.	.	52	13	6
<i>Artemisia hololeuca</i>	.	.	.	48	4	12
<i>Festuca cretacea</i>	.	.	.	18	1	1
<i>Asperula tephrocarpa</i>	.	.	.	62	55	48
<i>Plantago maritima</i> subsp. <i>ciliata</i>	.	.	.	16	.	6
All. <i>Euphorbio cretophilae</i>-<i>Thymion cretacei</i>						
<i>Euphorbia petrophila</i>	.	.	.	7	84	5
<i>Jurinea stoechadifolia</i>	.	.	.	1	73	7
<i>Helianthemum canum</i> aggr.	.	.	.	2	28	2
<i>Hedysarum grandiflorum</i>	.	.	1	2	32	10

ID clusters	1	2	3	4	5	6
Number of relevés	35	11	73	243	92	28
<i>Reseda lutea</i>	.	.	1	23	55	38
<i>Genista scythica</i>	.	.	.	.	20	1
<i>Linum tenuifolium</i>	.	.	.	5	25	4
<i>Onosma simplicissima</i>	3	.	11	14	49	38
<i>Leontodon crispus</i> aggr.	.	.	.	.	16	5
<i>Rhaponticoides talievii</i>	.	.	.	.	11	.
All. <i>Centaureo carbonatae</i>-<i>Koelerion talievii</i>						
<i>Carex humilis</i>	.	.	.	2	3	44
<i>Stipa pennata</i>	.	.	.	2	1	33
<i>Festuca valesiaca</i> aggr.	9	.	7	9	5	51
<i>Viola ambigua</i>	.	.	.	1	5	33
<i>Polygala sibirica</i>	.	.	.	6	1	33
<i>Bromopsis riparia</i>	.	.	.	2	8	35
<i>Stipa capillata</i>	3	.	3	4	15	44
<i>Bupleurum falcatum</i>	.	.	.	16	18	45
<i>Securigera varia</i>	.	.	.	4	9	31
<i>Koeleria talievii</i>	.	.	.	7	10	34
<i>Euphorbia seguieriana</i>	34	9	10	25	8	64
<i>Campanula sibirica</i>	.	.	.	5	35	44
<i>Stachys recta</i>	.	.	.	3	5	26
<i>Salvia nutans</i>	.	.	.	1	29	39
<i>Achillea millefolium</i> aggr.	.	.	.	.	.	16
<i>Astragalus austriacus</i>	.	.	1	1	1	20
<i>Medicago falcata</i>	3	.	1	5	7	29
<i>Scabiosa ochroleuca</i>	.	.	.	7	10	29
<i>Vincetoxicum hirundinaria</i>	.	.	.	2	21	31
<i>Androsace villosa</i>	.	.	.	5	.	19
<i>Polygala cretacea</i>	.	.	1	26	9	36
<i>Ajuga chamaepitys</i>	.	.	.	.	.	11
<i>Jurinea mollis</i> aggr.	.	.	.	5	5	20
<i>Euphorbia nicaeensis</i>	.	.	.	9	3	21
<i>Helianthemum nummularium</i>	.	.	.	.	2	13
Ord. <i>Anabasietaalia cretaceae</i>						
<i>Psathyrostachys juncea</i>	49	.	38	1	.	.
<i>Seseli glabratum</i>	14	82	89	.	.	.
Ord. <i>Thymo cretacei</i>-<i>Hyssopetalia cretacei</i>						
<i>Pimpinella tragioides</i>	.	.	4	86	83	58
<i>Cephalaria uralensis</i>	.	.	11	60	70	29
<i>Teucrium polium</i>	.	.	.	20	75	68
<i>Thymus calcareus</i>	.	.	.	70	87	83
<i>Gypsophila oligosperma</i>	.	.	.	57	71	66
Other taxa occurred in the communities						
<i>Thalictrum minus</i>	.	.	.	.	23	26
<i>Astragalus albicaulis</i>	.	.	.	23	13	32
<i>Poa compressa</i>	.	.	.	11	4	19
<i>Salvia verticillata</i>	.	.	.	3	11	18
<i>Erysimum diffusum</i>	6	.	3	3	15	23
<i>Helichrysum arenarium</i>	.	.	.	13	20	25
<i>Linum flavum</i> aggr.	.	.	.	37	47	40
<i>Linum hirsutum</i>	.	.	.	7	23	22
<i>Silene supina</i>	.	.	.	28	22	26
<i>Psephellus marschallianus</i> aggr.	34	.	36	5	34	34
<i>Thesium ramosum</i>	.	.	3	13	12	16
<i>Stipa lessingiana</i>	14	.	21	4	13	18

ID clusters	1	2	3	4	5	6
Number of relevés	35	11	73	243	92	28
<i>Artemisia lerchiana</i>	37	18	33	2	.	1
<i>Brassica elongata</i>	23	.	23	14	38	17
<i>Odontarrhena tortuosa</i> aggr.	6	.	33	2	38	9
<i>Genista tinctoria</i> aggr.	.	.	.	19	21	14
<i>Odontites luteus</i>	.	.	.	13	16	6
<i>Echinops ritro</i>	23	45	41	5	4	14
<i>Crambe tataria</i> aggr.	11	.	22	3	.	2
<i>Camphorosma monspeliaca</i>	11	.	15	.	.	.

Cluster 1

35 relevés (Table 1, Fig. 3, 4)

Diagnostic species: *Agropyron cristatum* aggr., *Allium inderiense*, *A. lineare*, *A. tulipifolium*, *Anabasis salsa*, *Androsace maxima*, *Artemisia lessingiana*, *A. semiarida*, *Asparagus inderiensis*, *Astragalus kustanaicus*, *A. temirensis*, *Bassia prostrata*, *Caragana balchaschensis*, *Eriosynaphe longifolia*, *Ferula caspica*, *Filago arvensis*, *Gagea bulbifera*, *Galatella tatarica*, *Isatis gymnocarpa*, *Ixiolirion tataricum*, *Klasea erucifolia*, *Krascheninnikovia ceratoides*, *Limonium meyeri*, *Linaria incompleta*, *Litwinowia tenuissima*, *Meniocus linifolius*, *Ornithogalum fischerianum*, *Palimbia turgaica*, *Poa bulbosa*, *Psathyrostachys juncea*, *Pseudosedum lievenii*, *Rindera tetraspis*, *Rochelia disperma* subsp. *retorta*, *Silene otites* aggr., *S. viscosa*, *Sterigmostemum caspicum*, *Stipa sareptana*, *Takhtajaniantha pusilla*, *Taraxacum* sect. *Macrocornuta*, *Tragopogon ruber*, *Tulipa biflora*, *T. scythica*, *T. suaveolens*, *T. sylvestris*.

Ecology and distribution: These communities occur at the feet of chalk hills, on alluvial fans of chalk bedrocks and chalk plateau peaks facing south, and south-western slopes with an inclination of 30–75° of the Sub-Ural Upland (Kazakhstan). They are apparently the latest stage of the chalk overgrowth and are transitional between *Festuco-Brometea* and *Anabasi-etalia cretaceae* order *nom. inv.* (Golovanov et al. 2021).

Cluster 2

11 relevés (Table 1, Fig. 3, 4)

Diagnostic species: *Anabasis cretacea*, *Artemisia terrae-albae*, *Astragalus medius*, *A. stenoceras*, *Atraphaxis replicata*, *A. spinosa*, *Convolvulus fruticosus*, *Ephedra distachya*, *Galium humifusum*, *Gypsophila diffusa*, *Lepidium meyeri*, *Limonium suffruticosum*, *Lomelosia isetensis*, *Matthiola tatarica*, *Nanophyton erinaceum*, *Onosma staminea*, *Rhammatophyllum pachyrhizum*, *Seseli glabratum*, *Silene antri-jovis*.

Ecology and distribution: These communities occur on chalk outcrops on slopes facing south to north and northwest in northern deserts and desert steppes in the aridic conditions of the Sub-Ural Upland (Kazakhstan and partly Russian Federation). They occupy scree slopes with an inclination of 30–75° (Golovanov et al. 2021).

Cluster 3

73 relevés (Table 1, Fig. 3,4)

Diagnostic species: *Archanthemis trotzkiana*, *Artemisia salsoloides*, *Astragalus tenuifolius*, *Atraphaxis decipiens*, *A. frutescens*, *Galium octonarium*, *Gypsophila altissima*, *G. rupestris*, *Hedysarum razoumovianum*, *H. tscherkassovae*, *Jurinea kirghisorum*, *Matthiola fragrans*, *Psathyrostachys juncea*, *Seseli glabratum*, *Zygophyllum pinnatum*.

Ecology and distribution: These communities occur on chalk uplands with gravel substrates or eroded and loose chalk outcrops on slopes facing mainly to the north of the Sub-Ural and partly Obshchy Syrt Uplands (Kazakhstan). They occupy scree slopes with an inclination of 15–60° (Golovanov et al. 2021).

Cluster 4

243 relevés (Table 1, Fig. 3, 4)

Diagnostic species: *Artemisia hololeuca*, *Asperula tephrocarpa*, *Cephalaria uralensis*, *Festuca cretacea*, *Hyssopus officinalis*, *Pimpinella tragioides*, *Plantago maritima* subsp. *ciliata*, *Scrophularia cretacea*.

Ecology and distribution: These communities occur on chalk outcrops at elevations of 70–140 m a.s.l. on scree slopes (15–65°) facing north to northwest on the right banks of river valleys. They occur on loose substrates caused by denudation and cover surface erosion. Relevés belonging to this cluster were recorded in the Central Russian Upland, Donetsk Ridge (Ukraine and Russian Federation). The layer of bryophytes and lichens is consisted of *Enchylium tenax*, *Geheebia fallax*, *Syntrichia ruralis*, *Weissia levieri*.

Cluster 5

92 relevés (Table 1, Fig. 3, 4)

Diagnostic species: *Cephalaria uralensis*, *Euphorbia petrophila*, *Genista scythica*, *Gypsophila oligosperma*, *Hedysarum grandiflorum*, *Helianthemum canum* aggr., *Jurinea stoechadifolia*, *Leontodon crispus* aggr., *Linum tenuifolium*, *Onosma simplicissima*, *Pimpinella tragioides*, *Reseda lutea*, *Rhaponticoides talievii*, *Teucrium polium*, *Thymus calcareus*.

Ecology and distribution: These communities occur on the lowest line of gentle chalk slopes (5–40°) facing south and southwest of river valleys. They indicate the final stage of overgrowing and transformation into pure grasslands. These communities occupy dense diluvium with carbonate and sulphate soil salinization and occur in the Central Russian Upland, Donetsk Ridge, and Eastern Don Ridge (Ukraine, Russian Federation).

Cluster 6

280 relevés (Table 1, Fig. 3, 4)

Diagnostic species: *Achillea millefolium* aggr., *Ajuga chamaepitys*, *Androsace villosa*, *Astragalus austriacus*, *Bromopsis riparia*, *Bupleurum falcatum*, *Campanula sibirica*, *Carex humilis*, *Euphorbia nicaeensis*, *E. seguieriana*, *Festuca valesiaca* aggr., *Gypsophila oligosperma*, *Helianthemum nummularium*, *Jurinea mollis* aggr., *Koeleria talievii*, *Medicago falcata*, *Polygala cretacea*, *P. sibirica*, *Salvia nutans*, *Scabiosa ochroleuca*, *Securigera varia*, *Stachys recta*, *Stipa capillata*, *S. pennata*, *Teucrium polium*, *Thymus calcareus*, *Vincetoxicum hirundinaria*, *Viola ambigua*.

Ecology and distribution: These communities occur on depths of chalk outcrops at elevations of 75–145 m a.s.l. on slopes facing north and northeast on which rendzic leptosols have accumulated. They occur in the Central Russian Upland, Donetsk Ridge (Ukraine and Russian Federation). The layer of bryophytes and lichens is consisted of *Barbula unguiculata*, *Ceratodon purpureus*, *Cladonia rei*, *Enchylium tenax*, *Homalothecium lutescens*, *Syntrichia ruralis*, *Trichostomum crispulum*, *Weissia longifolia*.

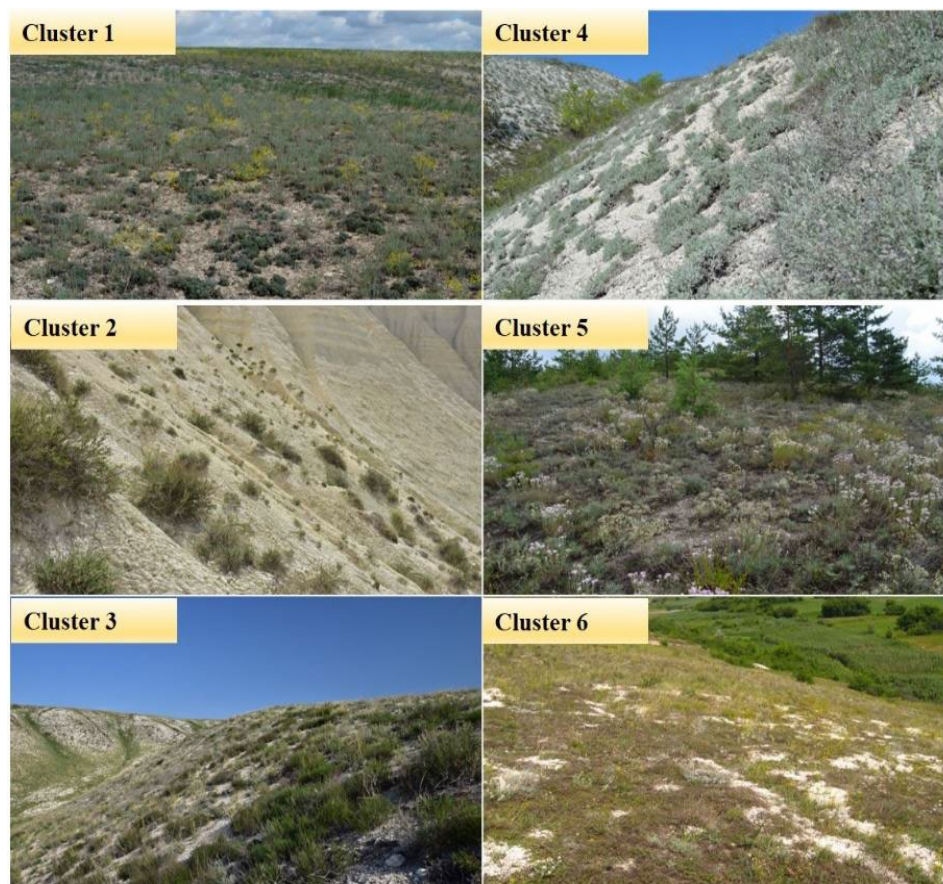


Fig. 3. Chalk rocky grasslands within the Eastern European Plain. Cluster 1 – *Anabasio cretaceae-Agropyron desertori* (Troitsk Chalk Hills, adjacent territories of Kazakhstan, 2019); Cluster 2 – *Sileno fruticosae-Nanophytion erinacei* (Aktolagai Ridge, Kazakhstan, 2019); Cluster 3 – *Anthemido troitzkianae-Artemision salsoloides* (Troitsk Chalk Hills, adjacent territories of Kazakhstan, 2019); Cluster 4 – *Sileno supinae-Artemision hololeucae* (Krasna valley, Svatove, Ukraine, 2014); Cluster 5 – *Euphorbio cretophilae-Thymion cretacei* (“Kreidova Flora” Nature Reserve, Ukraine, 2019); Cluster 6 – *Centaureo carbonatae-Koelerion talievii* (Krasna valley, Honcharivka, Ukraine, 2013) (Photos: cited from Golovanov et al. 2021 (1–3), O. Chusova (4–6)).

Abb. 3. Kalkfelsrasen in der osteuropäischen Tiefebene. Cluster 1 – *Anabasio cretaceae-Agropyron desertori* (Troizker Kreidehügel, angrenzende Gebiete Kasachstans, 2019); Cluster 2 – *Sileno fruticosae-Nanophytion erinacei* (Aktolagai-Rücken, Kasachstan, 2019); Cluster 3 – *Anthemido troitzkianae-Artemision salsoloides* (Troizker Kreidehügel, angrenzende Gebiete Kasachstans, 2019); Cluster 4 – *Sileno supinae-Artemision hololeucae* (Krasna-Tal, Svatove, Ukraine, 2014); Cluster 5 – *Euphorbio cretophilae-Thymion cretacei* (Naturschutzgebiet „Kreidova Flora“, Ukraine, 2019); Cluster 6 – *Centaureo carbonatae-Koelerion talievii* (Krasna-Tal, Honcharivka, Ukraine, 2013) Fotos: zitiert nach Golovanov et al. 2021 (1–3), O. Chusova (4–6)).

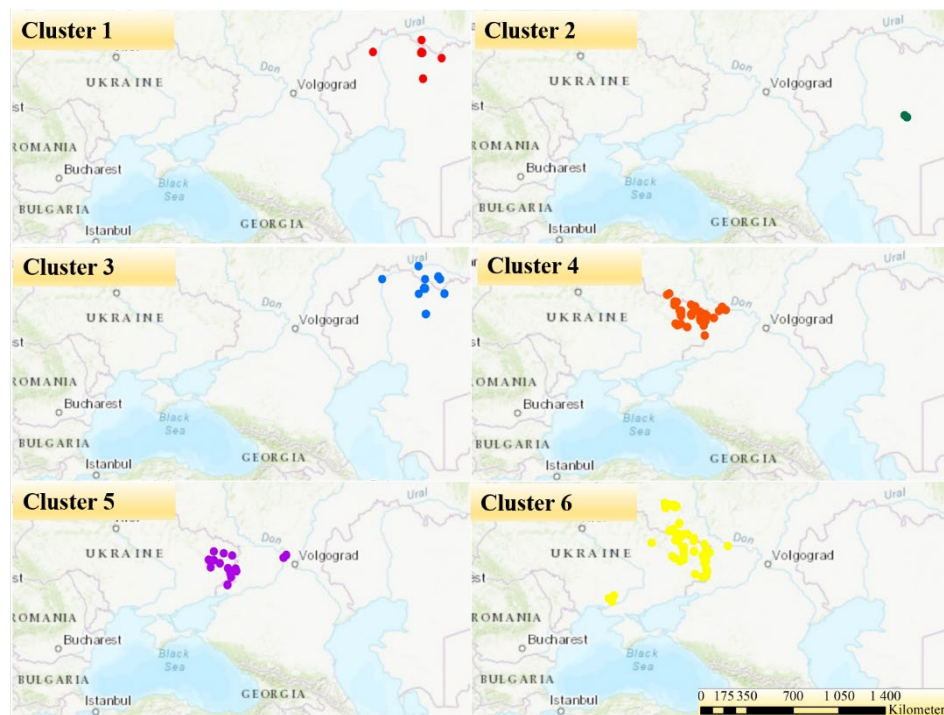


Fig. 4. Maps of the chalk rocky grasslands distribution (alliance level). Abbreviations are the same as in Figure 3.

Abb. 4. Verbreitungskarten der Kalkfelsrasen auf der Verbandsebene. Die Abkürzungen entsprechen denen in Abbildung 3.

Proposed syntaxonomic scheme

??? Class: *Helianthemo-Thymetea* Romashchenko et al. 1996

Order: *Anabasieta lia cretaceae* Golovanov et al. ex Vasheniak et al. 2024

Alliance: *Anabasio cretaceae-Agropyron desertori* Golovanov et al. 2021

Alliance: *Sileno fruticosae-Nanophytion erinacei* Golovanov et al. 2021

Alliance: *Anthemido trotziana-Artemision salsoloides* Golovanov et al. 2021

Order: *Thymo cretacei-Hyssopetalia cretaceae* Didukh ex Romashchenko et al. 1996

Alliance: *Sileno supinae-Artemision hololeuca* Didukh 1989 (*Artemisio hololeuca-Hyssopion cretacei* Romashchenko et al. 1996 nom. superfl.)

Alliance: *Euphorbio cretophilae-Thymion cretacei* Didukh ex Vasheniak et al. 2024

Transitional communities between *Helianthemo-Thymetea* and *Festuco-Brometea*

Alliance: *Centaureo carbonatae-Koelerion talievii* Romashchenko et al. 1996

4.2 Environmental differences within chalk rocky grasslands

Detrended correspondence analysis (DCA) shows the differentiation of chalk rocky grasslands (*Anabasietaia cretaceae*, *Thymo cretacei-Hyssopetalia cretaceae* orders) and other types of calcareous rocky grasslands (*Stipo pulcherrimae-Festucetalia pallentis* order) across the DCA2 axis where the vector of reaction lays within the DCA2 axis plane (Fig. 5). This differentiation is caused by variable calcareous bedrocks with high calcium ion content of chalks and lower in marls, dolomites and limestones, which affects the soil reaction.

Generally, a clear ecological distinction between chalk and other calcareous rocky grasslands can be observed; however, differentiation is also noticeable within the *Anabasietaia cretaceae* and *Thymo cretacei-Hyssopetalia cretaceae* group along the gradient of nitrogen, temperature and light. Therefore, we will focus more closely on the chalk rocky grasslands at the alliance level, using a broader range of ecological indicators based on Didukh's scales.

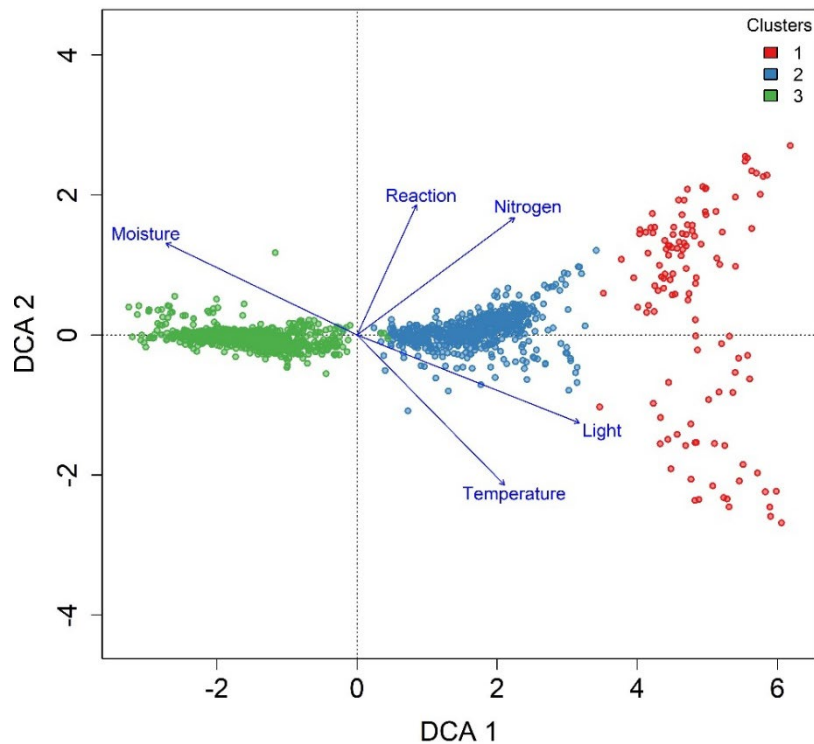


Fig. 5. DCA ordination of *Anabasietaia cretaceae* (1), *Thymo cretacei-Hyssopetalia cretaceae* (2), *Stipo pulcherrimae-Festucetalia pallentis* (3) based on EIVE 1.0 scales with passively projected mean indicator values.

Abb. 5. DCA-Ordination der *Anabasietaia cretaceae* (1), *Thymo cretacei-Hyssopetalia cretaceae* (2), *Stipo pulcherrimae-Festucetalia pallentis* (3) basierend auf EIVE 1.0-Skalen mit passiv projizierten mittleren Indikatorwerten.

According to Figure 6, chalk rocky grasslands differentiate across the DCA2 axis. The vectors of cryoregime, aeration, variation of damping and carbonate content indicators lie in the DCA1 axis plane. On the other hand, the vectors of nitrogen content and thermoregime lie in the DCA2 axis plane, where two groups (1–3 and 4–6) of chalk rocky grasslands are differentiated. Additionally, communities differ within the salt regime, continentality and light vectors, whereas communities of *Sileno fruticosae-Nanophytion erinacei* occupy an extreme position of ecological values. This differentiation is caused by the increasing continentality, surface illumination, considerable evaporation and further salinisation of the communities in Kazakhstan compared with Ukraine and Russian Federation. All communities vary within the reaction indicator vector, where communities of *Sileno fruticosae-Nanophytion erinacei* and *Anthemido trotzkianae-Artemision salsoloides* occupy the extreme position of the reaction indicator.

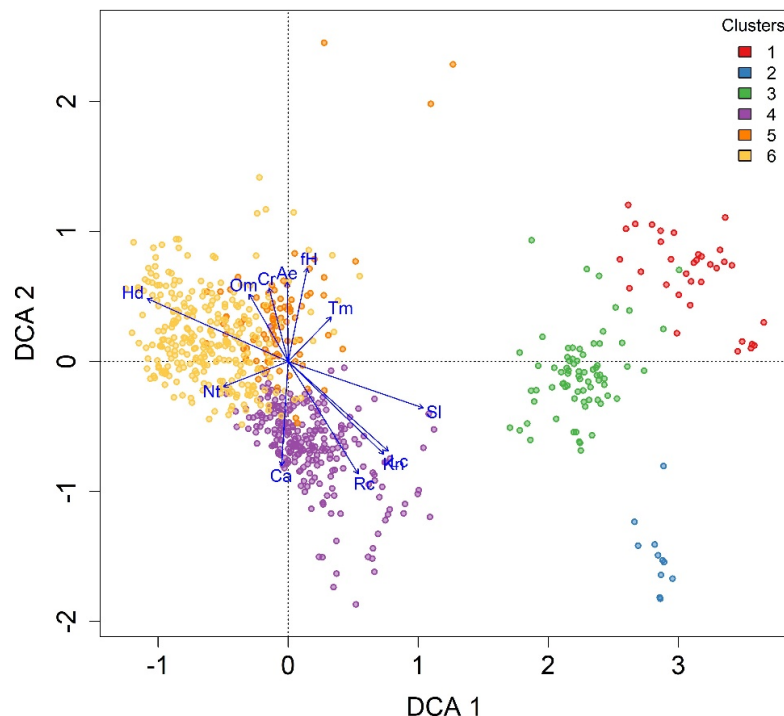


Fig. 6. DCA ordination of chalk rocky grasslands based on Didukh' scales with passively projected mean indicator values. **Abbreviations:** 1 – *Anabasio cretaceae-Agropyron desertori*, 2 – *Sileno fruticosae-Nanophytion erinacei*, 3 – *Anthemido trotzkianae-Artemision salsoloides*, 4 – *Sileno supinae-Artemision hololeucae*, 5 – *Euphorbio cretophilae-Thymion cretacei*, 6 – *Centaureo carbonatae-Koelerion talievii*. **Acronyms:** Hd – soil humidity, fH – variability of damping, Rc – soil acidity, Sl – total salt regime, Ca – carbonate content, Nt – nitrogen content, Ae – soil aeration, Tm – thermoregime, Om – humidity of microclimate, Kn – continentality, Cr – cryoregime, Lc – light in the community.

Abb. 6. DCA-Ordination von Kreidefelsrasen basierend auf Didukh-Skalen mit passiv projizierten mittleren Indikatorwerten. **Abkürzungen** der Verbände s.o. **Akronyme:** Hd – Bodenfeuchte, fH – Variabilität der Feuchtigkeit, Rc – Säuregehalt des Bodens, Sl – Gesamtsalzregime, Ca – Karbonatgehalt, Nt – Stickstoffgehalt, Ae – Bodenbelüftung, Tm – Thermoregime, Om – Feuchtigkeit des Mikroklimas, Kn – Kontinentalität, Cr – Kryoregime, Lc – Licht in der Gesellschaft.

To gain a better understanding of the structure and ecological characteristics of chalk rocky grasslands, which distinguish them from other types of grasslands, we carried out further analyses of life forms and ecological strategies to compare six syntaxa of chalk rocky grasslands with seven syntaxa of other types of calcareous rocky grasslands at the alliance level (see Supplement E4).

Life forms determine the structure, character of development, dynamics and resistance of coenoses to the influence of external factors; they, therefore, have significant diagnostic value. According to Figure 7, there is an apparent gap between chalk rocky grasslands and rocky grasslands of the *Stipo pulcherrimae-Festucetalia pallentis* order (Supplement E4). Communities belonging to the *Stipo pulcherrimae-Festucetalia pallentis* order have a significantly higher proportion of hemicryptophytes, the highest in the case of the *Seslerion rigidae* alliance (Fig. 7). In contrast, communities of the *Helianthemo-Thymetea* class have a much lower representation of hemicryptophytes but a much higher proportion of chamaephytes (Fig. 7), which is exceptionally high in the case of the alliances *Anthemido trotzkianae-Artemision salsoloides* and *Sileno fruticosae-Nanophytion erinacei*. An equal proportion of hemicryptophytes and chamaephytes is characteristic of the *Anabasio cretaceae-Agropyron desertori* and *Centaureo carbonatae-Koelerion talievii* alliances.

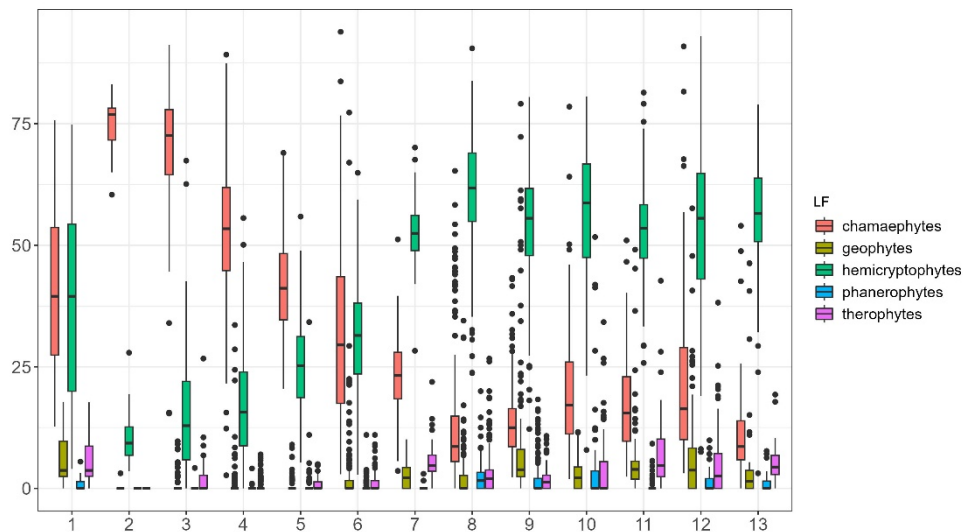


Fig. 7. Life forms of the distinguished thirteen clusters that represent chalk rocky grasslands compared with other types of calcareous grasslands (alliance level).

Abbreviations: 1 – *Anabasio cretaceae-Agropyron desertori*; 2 – *Sileno fruticosae-Nanophytion erinacei*; 3 – *Anthemido trotzkianae-Artemision salsoloides*; 4 – *Sileno supinae-Artemision hololeuca*; 5 – *Euphorbio cretophilae-Thymion cretacei*; 6 – *Centaureo carbonatae-Koelerion talievii*; 7 – *Seseli leucospermum-Fumana procumbens* grasslands; 8 – *Seslerion rigidae*; 9 – *Sesleria caerulea-Leontodon incanus* grasslands; 10 – *Asplenio septentrionalis-Festucion pallentis*; 11 – *Bromo pannonici-Festucion pallentis*; 12 – *Galio campanulati-Poion versicoloris*; 13 – *Linum catharticum-Galium cracoviense* grasslands.

Abb. 7. Lebensformen der dreizehn unterschiedenen Cluster, die die Kalkfelsrasen im Vergleich mit anderen Typen von Kalkmagerrasen darstellen (Verbandsebene). Abkürzungen der Syntaxa s. englische Abbildungsunterschrift.

According to the ecological strategies of Grime, chalk rocky grasslands are located in the fields (S, SC) close to S (> 50%), C < 50% and R < 75% with the dominance of stress-tolerant and stress-tolerant/competitor species. On the other hand, the calcareous rocky grasslands of *Stipo pulcherrimae-Festucetalia pallentis* are located within the CS field (Fig. 8).

SPE habitats occur at reach conditions, absorb and transform resources and energy and develop within the successional range. When the “centres of mass” approach the C peak, the mentioned parameters increase (Didukh 2023). FLE habitats are characterized by high lability, the stability of which is due to rapid reproduction after the destruction and high species flexibility caused by fluctuations characterised by resource and energy accumulation and rapid transmission (Didukh 2023). According to Figur 8, “centres of mass” do not occur in the R field, and fluctuations are not characteristic of FLE habitats.

PSE habitats occur within extreme conditions caused by maximum and minimum indices of factors that limit resource consumption. These communities are stable in relation to environmental impact, highly resistant, with limited successional changes, and exist indefinitely with weakly expressed development stages. PSE habitats develop slowly and express themselves in speciation processes due to changes in the morphological and anatomical features of organs as a specialization of adaptation mechanisms to external conditions of existence (Didukh 2023). When the “centres of mass” of chalk rocky grasslands approach the S peak, the mentioned parameters are characteristic, and chalk rocky grasslands differ from pure steppes and reflect high conservatism. Notably, the *Anabasio cretaceae-Agropyron desertori*, *Sileno fruticulosae-Nanophytion erinacei*, and *Anthemido trotzkianae-Artemision salsoloides* communities are the closest to the PSE field and could be characterized as the persistent stable ecosystems (Fig. 8). Thus, neither successions nor fluctuating processes are distinct, which is essential to consider in developing management measures for their conservation.

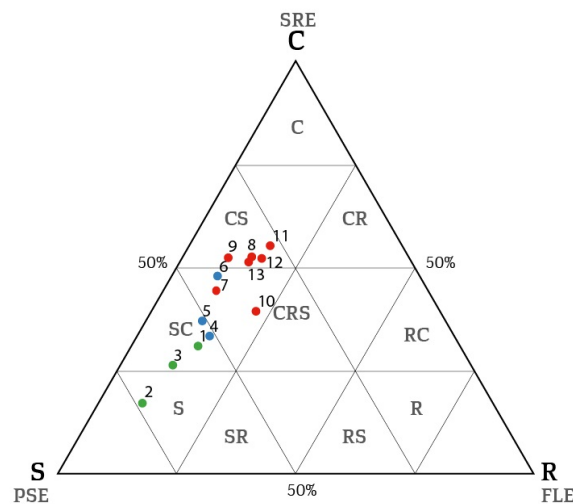


Fig. 8. The ecological strategy of the distinguished thirteen clusters that represent chalk rocky grasslands compared with other types of calcareous grasslands (alliance level). Abbreviations are the same as in the Figure 6.

Abb. 8. Die ökologische Strategie der dreizehn unterschiedenen Cluster, die Kalkfelsrasen im Vergleich zu anderen Arten von Kalkmagerrasen repräsentieren (Verbandsebene). Die Abkürzungen sind dieselben wie in Abbildung 6.

4.3 Syntaxonomical revision

Continental arid steppe zone that unifies meso-xerophytic grasslands on deep calcareous soils of Western and Central Europe that are classified within the *Festuco-Brometea* Br.-Bl. et Tx. ex Soó 1947. When conditions become more arid, European steppes give way to the semi-desert and desert steppe vegetation within the continental arid semidesert zone that syntaxonomically classified as the *Artemisietea lerchianae* Golub 1994 (Mucina et al. 2016, 2024). However, zonal vegetation of *Festuco-Brometea* and *Artemisietea lerchianae* occur on deep soils of different genesis where the correlation of edaphic and climate environmental parameters is significant (Didukh 2023). What about the cretaceous vegetation that occurs on shallow soils on chalk outcrops if they are variations of zonal vegetation?

Chalk rocky vegetation is floristically and ecologically different from the zonal steppe vegetation of *Festuco-Brometea* and *Artemisietea lerchianae* and was considered within two classes: *Helianthemo-Thymetea* and *Anabasietaea cretaceae nom. inv.* (Romashchenko et al. 1996, Didukh et al. 2018, Golovanov et al. 2021).

The *Thymo cretacei-Hyssopetalia cretacei* order, designated by Didukh (1989) as *Thymo-Hyssopetalia* in the original diagnosis, has the nomenclature type of the *Euphorbio cretophilae-Thymion* alliance. On the other hand, the *Euhorbio cretaceae-Thymion cretaceae* alliance was probably mistakenly provided in the original diagnosis (Art. 41a), rather than the *Euphorbio cretophilae-Thymion cretaceae* alliance, and it has the nomenclature type of the *Euphorbio cretaceae-Jurinetum brachycephalae* association (Didukh 1989). The name of the *Euphorbio cretophilae-Jurinetum brachycephalae* association is invalid (Art. 3f) since there is “*Euphorbia cretacea*” in the original diagnosis (Table 2, p. 19 in Didukh 1989); however, these taxa does not exist since it is not listed in Euro+Med PlantBase (<https://www.emplantbase.org/>). According to Art. 3f (Theurillat et al. 2021), the *Euphorbio cretophilae-Thymion* alliance and *Thymo-Hyssopetalia* order are invalidly published due to invalidly published *Euphorbio cretophilae-Jurinetum brachycephalae* association.

Romashchenko et al. (1996) later designated the illegitimate *Artemisio hololeucae-Hyssopion cretacei* Romashchenko et al. 1996 as a type of the *Thymo cretacei-Hyssopetalia cretacei* order and *de facto* validated the *Thymo-Hyssopetalia* order. The *Artemisio hololeucae-Hyssopion cretaceae* alliance is validly published, and the nomenclature type of the alliance is the *Artemisio hololeucae-Polygaletum cretaceae* association. Romashchenko et al. (1996) designated *Artemisio hololeucae-Polygaletum cretaceae* Didukh 1989 as the nomenclatural type of this alliance. However, Romashchenko et al. (1996) further indicated that this alliance is identical to *Sileno supinae-Artemision hololeucae* Didukh 1989. This means that *Artemisio hololeucae-Hyssopion cretaceae* Romashchenko et al. 1996 is “superfluous” (Art. 29c), hence the illegitimate name of *Sileno supinae-Artemision hololeucae* Didukh 1989. The *Sileno supinae-Artemision hololeucae* alliance is validly published by Didukh (1989) and designated the valid *Sileno supinae-Matthioletum fragrantis* association. The nomenclature type of the *Thymo cretacei-Hyssopetalia cretacei* order is, therefore, the *Sileno supinae-Artemision hololeucae* alliance.

Regarding the class *Anabasietaea cretaceae* and order *Anabasietaalia cretaceae*, the nomenclature types of high-rank syntaxa have been provided mistakenly and invalidly published (Art.8) (Theurillat et al. 2021). The nomenclature type of the order *Anabasietaalia cretaceae* and class *Anabasietaea cretaceae* is the *Anthemido trotzkianae-Artemisietum salsoloidis* association, despite the selected names of the alliance and order, respectively (Golovanov et al. 2021).

Regarding the *Helianthemo-Thymetea*, the nomenclature type of the class is the *Thymo cretacei-Hyssopetalia cretacei* order (Art. 8). Romashchenko et al. (1996) have validated the class in their paper, although they have not explicitly cited correct name “*Thymo cretacei-Hyssopetalia cretacei* Didukh ex Romashchenko et al. 1996” according to the Code (Theurillat et al. 2021). Mucina et al. (2016) do not consider this class in its own right and assign chalk rocky grasslands within the *Festuco-Brometea* class. However, chalk rocky grasslands sharply differentiate from the closer type of vegetation that occurs on calcareous outcrops (*Stipo pulcherrimae-Festucetalia pallentis*) (Fig. 2) and have their own diagnostic species (e.g. *Artemisia salsoloides*, *Cephalaria uralensis*, *Pimpinella tragium*). On the other hand, there are many species (e.g. *Asperula cynanchica*, *Campanula sibirica*, *Festuca valesiaca* aggr.) with high constancy that are considered diagnostic for *Festuco-Brometea*; therefore, we have preliminarily assigned chalk rocky grasslands within *Helianthemo-Thymetea* class” (Kuzemko & Khodosovtsev 2022) with question marks “???”.

4.4 Validation of the chalk rocky grasslands syntaxa

We propose to designate the nomenclature type of the *Euphorbio cretophilae-Jurinetum brachycephalae* association from Romashchenko et al. (1996) and validate the name of the association (Art. 2b):

Association *Euphorbio cretophilae-Jurinetum brachycephalae* Didukh ex Vasheniak et al. 2024

Neotypus hoc loco: Romashchenko et al. (1996), page 57, table 3, relevé 24.

Diagnostic species: according to Romashchenko et al. (1996): *Centaurea lavrenkoana*, *Cytisus ruthenicus*, *Genista tinctoria*, *Helichrysum arenarium*, *Linum austriacum*, *Minuartia setacea*, *Odontarrhena tortuosa*.

The name of the *Euphorbio cretophilae-Jurinetum brachycephalae* association is assigned by the authors (Romashchenko et al. 1996) as the nomenclature type of the alliance *Euphorbio cretophilae-Thymion cretacei* alliance. We propose validating the alliance name according to the association's validation (Art. 2b, 8).

Alliance *Euphorbio cretophilae-Thymion cretacei* Didukh ex Vasheniak et al. 2024

Neotypus hoc loco: *Euphorbio cretophilae-Jurinetum brachycephalae* Didukh ex Vasheniak et al. 2024.

Diagnostic species: according to Romashchenko et al. (1996): *Artemisia salsoloides*, *Brassica elongata* subsp. *pinnatifida*, *Euphorbia petrophila*, *Jurinea stoechadifolia*.

In addition, we propose the designation of the validly published *Anthemido troztkianae-Artemision salsoloidis* alliance (Golovanov et al. 2021) as the nomenclature type of the *Anabasietales cretaceae* order (Art. 2b, 8).

Order *Anabasietales cretaceae* Golovanov et al. ex Vasheniak et al. 2024

Lectotypus hoc loco: *Anthemido troztkianae-Artemision salsoloidis* Golovanov et al. 2021.

Diagnostic species: according to Golovanov et al. (2021): *Anabasis cretacea*, *Anthemis troztkiana*, *Artemisia salsoloides*, *Atraphaxis decipiens*, *Crambe aspera*, *Echinops meyeri*, *Jurinea kirghisorum*, *Hedysarum tscherkassovae*, *Lepidium meyeri*, *Limonium cretaceum*, *Linaria cretacea*, *Matthiola fragrans*, *Nanophyton erinaceum*, *Seseli glabratum*, *Zygophyllum pinnatum*.

4.5 Distribution of chalk rocky grasslands within the Eastern European Plain

Chalk rocky grasslands sporadically spread within the steppe zone of the arid climate, on the other hand, occur in the south of the forest-steppe and the north of the semidesert zone and should be considered to be azonal-zonal vegetation (Lavrenko 1961) that differs from plain zonal meso-xerophytic grasslands on deep calcareous soils of the *Festuco-Brometea* and semidesert grasslands of *Artemisieta lerchianae* (Mucina et al. 2016, 2024).

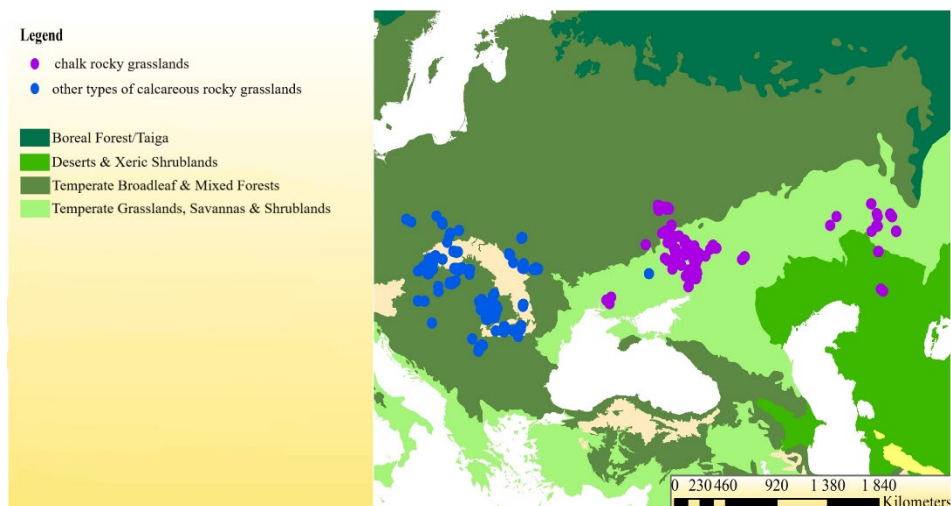


Fig. 9. Biomes and Ecoregions map of the chalk rocky grasslands compared with other types of calcareous rocky grasslands.

Abb. 9. Karte der Biome und Ökoregionen der untersuchten Kalkfelsrasen im Vergleich zu anderen Typen von Kalkfelsrasen.

On the one hand, *Centaureo carbonatae-Koelerion talievii* communities occur in steppe zone, which could be explained by the transitional nature of the alliance, which has a lot of similar features to the *Festucetalia valesiacae* order (*Festuco-Brometea* class) (Roma-shchenko et al. 1996, Poluyanov 2009, Poluyanov and Averinova 2012, Mucina et al. 2016, Didukh et al. 2018). Three plots of *Potentillo arenariae-Linon czernjajevii* Krasova et Smetana 1999, which were initially considered within the *Festucetalia valesiacae* order (Krasova and Smetana 1999), *Cephalario uralensis-Jurineetalia stoechadifolii* order (Vynokurov 2021) and more recently as the *Stipo pulcherrimae-Festucetalia pallentis* order (Mucina et al. 2016, Didukh et al. 2021) and have unclear syntaxonomical affiliation, are floristically very similar to *Centaureo carbonatae-Koelerion talievii* communities. Outliers of chalk rocky grasslands that occur in Right-bank Ukraine (Kherson region), according to the map (Fig. 9), cannot be considered within the *Thymo cretacei-Hyssopetalia cretaceae* order.

On the other hand, *Sileno fruticosae-Nanophytion erinacei* and *Anthemido troztkianae-Artemision salsoloides* communities occur in the arid semidesert zone and are deeply spread within it (Golovanov et al. 2021; Fig. 9). We presume that chalk rocky grasslands could be azonal vegetation, which strongly correlates with the edaphic parameters and is characterised by endemic races or species that are sporadically spread on chalk outcrops within the Eastern

European Plain (Klokov and Dobrochayeva 1974, Tzvelev et al. 1996, Chusova and Didukh 2014). It should be noted that the *Anabasietaia cretaceae* and *Thymo cretacei-Hyssopetalia cretaceae* orders are separated by the Volga Lowland; however, the high terraces of the Don River that occur in the Volga-Don interfluvium, are habitats of *Euphorbia cretophilae-Thymion cretacei* communities (Koroliuk et al. 2021; Fig. 9). Thus, in order to fill the gap between the two orders and find transitional communities within the Volga-Don interfluvium, we aim further to conduct a supraregional analysis of chalk rocky grasslands.

Edaphic and orographic parameters (uplands with chalk outcrops and undeveloped soils on the surface) vary within an arid climate with low precipitation and high evaporation (Didukh 2023). As the continentality indices increase and the indicators of the ombroregime decrease, chalk rocky grasslands change from the transitional *Centaureo carbonatae-Koelerion talievii* alliance to typical *Thymo cretacei-Hyssopetalia cretacei*, *Anabasietaia cretaceae* communities. According to the phytoindication results, continentality and ombroregime correlate with decreased soil humidity, nitrogen content and increased acidity and salt regimes (Fig. 5B). It should be noted that loose and disintegrated substrates of chalk rocky grasslands with a lack of soils differ from the wet, dense substrates of the *Stipo pulcherrimae-Festucetalia pallentis* communities, sub-Mediterranean and supra-oreomediterranean belts of *Drypidetea spinosae*, rocky communities of *Sedo-Scleranthetea* annuals and succulents and sandy grasslands of *Koelerio-Corynephoretea* (Mucina et al. 2016).

5. Conclusions

We provided a syntaxonomy of chalk rocky grasslands in the Eastern European Plain concluding that all communities belong to the *Thymo cretacei-Hyssopetalia cretacei* and *Anabasietaia cretaceae* orders. They are distinct from other steppe communities and belong to the azonal class *Helianthemo-Thymetea* with a high proportion of chamaephytes. Chalk rocky grasslands have a high presence of stress-tolerant/competitor species (SC) characterised as PSE habitats that occur under extreme conditions. A key aspect that distinguishes *Helianthemo-Thymetea* communities from typical steppe communities is that they form on rocky substrates in the absence of soil, which means that successional processes do not occur here. We consider such communities to be “edaphic-climax”.

The main environmental factors causing the differentiation of the *Helianthemo-Thymetea* communities are the soil reaction, carbonate content, aeration, continentality, cryoregime and light. The *Helianthemo-Thymetea* communities differ from other calcareous rocky grasslands along the moisture, light and nitrogen gradients. The distribution of chalk rocky grasslands is concentrated in the continental arid steppe zone, but they may also be found in the continental arid semidesert and nemoral zone, where climate (temperature, precipitation and continentality) and edaphic conditions (carbonate content, salt regime and soil moisture) vary significantly within three biogeographical zones.

Erweiterte deutsche Zusammenfassung

Einleitung – Kalkfelsrasen, die von bestimmten endemischen Chamaephyten und Hemikryptophyten mit sternförmiger und spinnennetzartiger Behaarung dominiert werden, kommen auf den Kalkfelsen der Flusstäler Osteuropas vor. Sie besiedeln schwach entwickelte Kalkskelettböden (Syrosemi, flachgründige Rendzinen) und sind stark vom hohen Karbonatgehalt des Kalkgrundgesteins abhängig. Kalkfelsrasen sind derzeit gefährdet und für den Schutz der Artenvielfalt von großer

Bedeutung. Die Hauptbedrohungen sind Aufforstung, Terrassierung der Hänge und der Abbau von Kreide und Mergel (Kuzemko 2022). Seit der russischen Aggression im Jahr 2014 sind neue Bedrohungen entstanden, die die gefährdeten Kalkfelsenrasen beeinflussen: direktes Verschwinden der Vegetationsdecke aufgrund intensiver militärischer Aktivitäten, aufgegebene landwirtschaftliche Flächen in der Nähe der Hänge von Kalkfelsen und Ausbreitung invasiver Arten. Sie gelten aber gemäß der Berner Konvention als EUNIS-Habitattyp E1.13 (kontinentale trockene, felsige Steppengrasländer und Zwerggestrüpp auf Kalkfelsen) als für den Naturschutz in Europa bedeutsam (Schaminée et al. 2016). Das Hauptziel der vorliegenden Studie war es, die Klassifizierung der Kreidefelsenvegetation in der osteuropäischen Tiefebene anhand eines großen internationalen Datensatzes zu überarbeiten und das Wissen über ihre Syntaxonomie, Ökologie und Verbreitung zu vervollständigen.

Untersuchungsgebiet – Wir haben Grasland auf Kalkfelsen in der Trockenzone der osteuropäischen Tiefebene untersucht. Insbesondere haben wir uns auf zwei unterschiedliche Regionen konzentriert: (1) die Mittellrussische Platte und (2) das Sub-Urale Plateau über einen Klimagradienten hinweg, der die Verteilung und Zusammensetzung der Vegetationstypen innerhalb des Untersuchungsgebiets, in dem Kalkfelsen vorkommen, erheblich beeinflusst. Unser Untersuchungsgebiet umfasst ein breites Klimaspektrum, das von den milden kontinentalen Bedingungen der Ukraine bis zu den rein kontinentalen Bedingungen Kasachstans reicht. Dieser Klimagradient beeinflusst die Verteilung und Zusammensetzung der Vegetationstypen innerhalb des Untersuchungsgebiets erheblich.

Material und Methoden – Um eine Auswahl der Zielpflanzengesellschaften zu erhalten, die der Klasse *Helianthemo-Thymetea* entsprechen, klassifizierten wir den ursprünglichen Datensatz von 3171 Vegetationsaufnahmen mithilfe von Expertensystemen (Willner et al. 2017, Chytrý et al. 2020). Wir verwendeten den modifizierten TWINSpan-Algorithmus (Hill 1979, Roleček et al. 2009): drei Pseudospezies-Schnittwerte – 0 %, 5 % und 25 %, Mindestgruppengröße für die Unterteilung – 10 Aufnahmen, und Whittakers Beta-Koeffizient als Maß für die interne Clusterheterogenität. Die diagnostischen Arten für einzelne Cluster wurden anhand des *phi*-Koeffizienten (Treueschwelle bei *phi* = 0,30) geschätzt, getestet mit dem exakten Test von Fisher (*p* < 0,05) (Chytrý et al. 2002), wobei die Größen aller Gruppen auf gleiche Größe standardisiert wurden (Tichý & Chytrý 2006). Um standörtliche Unterschiede zwischen den Clustern zu untersuchen, verwendeten wir Indikatorwerte vom Ellenberg-Typ und Didukhs Skalen (Didukh 2011, Dengler et al. 2023).

Ergebnisse und Diskussion – Wir haben den Datensatz mit 1643 Vegetationsaufnahmen mit dem modifizierten TWINSpan-Algorithmus klassifiziert. Die TWINSpan-Klassifizierung ergab 13 Cluster. Wir haben 7 Cluster entfernt, die als *Stipo pulcherrimae-Festucetalia pallentis* interpretiert wurden, und uns auf die verbleibenden sechs Cluster als die Verbände *Anabasio cretaceae-Agropyrion desertori*, *Sileno fruticosae-Nanophytion erinacei*, *Anthemido trozkianae-Artemision salsoloides*, *Sileno supinae-Artemision hololeucae*, *Euphorbio cretophilae-Thymion cretacei*, *Centaureo carbonatae-Koelerion talievii* konzentriert. Die Gesellschaften unterscheiden sich deutlich entlang der Gradienten der Klima- und Bodenparameter. Wir haben 7 Cluster zurückgegeben, die als *Stipo pulcherrimae-Festucetalia pallentis* interpretiert wurden und von der Hauptanalyse zum Vergleich von Umweltmerkmalen, Lebensformen und Verteilung der Gesellschaften ausgeschlossen wurden. Gesellschaften der Ordnung *Stipo pulcherrimae-Festucetalia pallentis* weisen einen deutlich höheren Anteil an Hemikryptophyten auf, den höchsten im Fall des Verbandes *Seslerion rigidae*. Im Gegensatz dazu sind in Gesellschaften der Klasse *Helianthemo-Thymetea* Hemikryptophyten viel weniger vertreten, aber der Anteil an Chamaephyten ist wesentlich höher. Wir haben einen aktuellen syntaxonomischen Überblick über die Gesellschaften der Grasländer auf Kalkfelsen in der osteuropäischen Tiefebene gegeben (vgl. Romashchenko et al. 1996, Didukh et al. 2018, Golovanov et al. 2021). Es wurden Übergangsgesellschaften vom *Centaureo carbonatae-Koelerion talievii* beobachtet, die Fragen zu ihrer Klassifizierung aufwerfen. Wir schlagen vor, einige von Didukh (1989), Romashchenko et al. (1996) und Golovanov et al. (2021) ungültig veröffentlichte Syntaxa zu validieren. Die Verbreitung der Kalkfels-Grasländer konzentriert sich auf die kontinentale aride Steppenzone; sie könnten jedoch auch in der kontinentalen ariden Halbwüste und der nemoralen Zone

zu finden sein, wo Klima (Temperatur, Niederschlag und Kontinentalität) und edaphische Bedingungen (Karbonatgehalt, Salzregime und Bodenfeuchtigkeit) innerhalb dreier biogeografischer Zonen erheblich variieren. Wir vermuten, dass es sich bei den Kalkfelsrasen um eine azonale Vegetation handeln könnte, die stark mit den edaphischen Parametern korreliert und durch endemische Rassen oder Arten gekennzeichnet ist, die sporadisch auf Kalkaufschlüssen in der osteuropäischen Tiefebene verbreitet sind (Klokov und Dobrochayeva 1974, Tzvelev et al. 1996, Chusova und Didukh 2014).

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Authors contributions

Y.D. conceived the research project. Y.D. and O.C. provided data on chalk rocky grasslands within the study area. I.V. performed the numerical analyses and wrote the manuscript, supervised by Y.D. I.V. prepared the electronic supplements. O.C. prepared the DCA and box-and-whiskers plots. I.V. and O.C. prepared the figures for publication. All the authors revised the manuscript and agreed on the final version for publication.

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Supplements

Additional supporting information may be found in the online version of this article.

Zusätzliche unterstützende Information ist in der Online-Version dieses Artikels zu finden.

Supplement E1. List of aggregate species and definition of their combination.

Anhang E1. Liste der aggregierten Arten und Definition ihrer Kombination.

Supplement E2. Life forms, Grime's strategy for the species list used in the analysis.

Anhang E2. Lebensformen, Grime's Strategie für die Artenliste, die in der Analyse genutzt wurden.

Supplement E3. Algorithm of the Grime's triangle construction.

Anhang E3. Algorithmus zur Konstruktion des Grime-Dreiecks.

Supplement E4. Full synoptic table showing the percentage constancy and fidelity of species of the six clusters.

Anhang E4. Vollständige synoptische Tabelle mit der prozentualen Stetigkeit und Treue der Arten der sechs Cluster.

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Supplement E1. List of aggregate species and definition of their combination.

Anhang E1. Liste der aggregierten Arten und Definition ihrer Kombination.

Aggregates and valid names	Initial species data	Euro+Med	Source for aggregate definition	Synonyms
Achillea millefolium aggr.	Achillea collina	Achillea collina	Euro+Med (2024)	
Achillea millefolium aggr.	Achillea distans	Achillea distans	Euro+Med (2024)	
Achillea millefolium aggr.	Achillea millefolium	Achillea millefolium	Euro+Med (2024)	
Achillea millefolium aggr.	Achillea millefolium subsp. millefolium	Achillea millefolium subsp. millefolium	Euro+Med (2024)	
Achillea millefolium aggr.	Achillea millefolium subsp. pannonica	Achillea millefolium subsp. pannonica	Euro+Med (2024)	
Achillea millefolium aggr.	Achillea pannonica	Achillea pannonica	Euro+Med (2024)	
Achillea millefolium aggr.	Achillea setacea	Achillea setacea	Euro+Med (2024)	
Achillea millefolium aggr.	Achillea stricta	Achillea distans subsp. stricta	Euro+Med (2024)	
Agrimonia eupatoria aggr.	Agrimonia eupatoria	Agrimonia eupatoria	Ad hoc	
Agrimonia eupatoria aggr.	Agrimonia procera	Agrimonia procera	Ad hoc	Agrimonia eupatoria subsp. procera (Wallr.) Arrh. ex Fr
Agropyron cristatum aggr.	Agropyron cristatum	Agropyron cristatum	Ad hoc, based on Mosyakin & Fedoronchuk (1999)	Agropyron cristatum
Agropyron cristatum aggr.	Agropyron cristatum subsp. cristatum	Agropyron cristatum	Ad hoc, based on Mosyakin & Fedoronchuk (1999)	Agropyron cristatum
Agropyron cristatum aggr.	Agropyron desertorum	Agropyron desertorum	Ad hoc, based on Mosyakin & Fedoronchuk (1999)	Agropyron cristatum subsp. desertorum
Agropyron cristatum aggr.	Agropyron pectinatum	Agropyron cristatum subsp. pectinatum	Ad hoc, based on Mosyakin & Fedoronchuk (1999)	Agropyron cristatum subsp. pectinatum
Agropyron cristatum aggr.	Agropyron ponticum	Agropyron cristatum subsp. ponticum	Ad hoc, based on Mosyakin & Fedoronchuk (1999)	Agropyron cristatum subsp. ponticum
Allium angulosum aggr.	Allium angulosum	Allium angulosum	Landolt et al. (2010)	
Allium angulosum aggr.	Allium flavescens	Allium flavescens	Landolt et al. (2010)	
Allium angulosum aggr.	Allium montanum	Allium lustranicum	Landolt et al. (2010)	
Allium angulosum aggr.	Allium senescens	Allium senescens	Landolt et al. (2010)	
Allium angulosum aggr.	Allium senescens subsp. montanum	Allium senescens	Landolt et al. (2010)	
Allium paniculatum aggr.	Allium fuscum	Allium fuscum	Ad hoc	Allium paniculatum subsp. fuscum
Allium paniculatum aggr.	Allium paniculatum	Allium paniculatum	Ad hoc	Allium paniculatum
Allium paniculatum aggr.	Allium rupestre	Allium rupestre	Ad hoc	Allium paniculatum subsp. rupestre
Alyssum montanum aggr.	Alyssum gmelinii	Alyssum gmelinii	Landolt et al. (2010)	
Alyssum montanum aggr.	Alyssum montanum	Alyssum montanum	Landolt et al. (2010)	
Alyssum montanum aggr.	Alyssum montanum subsp. gmelinii	Alyssum montanum	Landolt et al. (2010)	
Alyssum montanum aggr.	Alyssum montanum subsp. montanum	Alyssum montanum	Landolt et al. (2010)	
Alyssum montanum aggr.	Alyssum repens	Alyssum repens	Landolt et al. (2010)	
Alyssum montanum aggr.	Alyssum rostratum	Alyssum rostratum	Landolt et al. (2010)	
Alyssum montanum aggr.	Alyssum trichostachyum	Alyssum trichostachyum	Landolt et al. (2010)	
Anthyllis vulneraria aggr.	Anthyllis biebersteiniana	Anthyllis vulneraria subsp. pulchella	Mosyakin & Fedoronchuk (1999)	
Anthyllis vulneraria aggr.	Anthyllis taurica	Anthyllis vulneraria subsp. boissieri	Mosyakin & Fedoronchuk (1999)	
Anthyllis vulneraria aggr.	Anthyllis vulneraria	Anthyllis vulneraria	Mosyakin & Fedoronchuk (1999)	
Anthyllis vulneraria aggr.	Anthyllis vulneraria subsp. alpestris	Anthyllis vulneraria subsp. alpestris	Mosyakin & Fedoronchuk (1999)	
Anthyllis vulneraria aggr.	Anthyllis vulneraria subsp. polyphylla	Anthyllis vulneraria subsp. polyphylla	Mosyakin & Fedoronchuk (1999)	
Anthyllis vulneraria aggr.	Anthyllis vulneraria subsp. vulneraria	Anthyllis vulneraria subsp. vulneraria	Mosyakin & Fedoronchuk (1999)	
Arenaria serpyllifolia aggr.	Arenaria leptoclados	Arenaria leptoclados	Ehrendorfer (1973)	
Arenaria serpyllifolia aggr.	Arenaria serpyllifolia	Arenaria serpyllifolia	Ehrendorfer (1973)	
Arenaria serpyllifolia aggr.	Arenaria serpyllifolia subsp. serpyllifolia	Arenaria serpyllifolia subsp. serpyllifolia	Ehrendorfer (1973)	
Arenaria serpyllifolia aggr.	Arenaria uraleensis	Arenaria leptoclados	Ehrendorfer (1973)	
Astragalus vesicarius aggr.	Astragalus albidus	Astragalus vesicarius L. subsp. vesicarius	Ad hoc	Astragalus vesicarius L. subsp. vesicarius
Astragalus vesicarius aggr.	Astragalus ciccr	Astragalus ciccr	Ad hoc	Astragalus vesicarius
Astragalus vesicarius aggr.	Astragalus vesicarius	Astragalus vesicarius	Ad hoc	Astragalus vesicarius
Campanula rotundifolia aggr.	Campanula kladniana	Campanula kladniana	Landolt et al. (2010)	
Campanula rotundifolia aggr.	Campanula moravica	Campanula rotundifolia	Landolt et al. (2010)	
Campanula rotundifolia aggr.	Campanula moravica subsp. moravica	Campanula rotundifolia	Landolt et al. (2010)	
Campanula rotundifolia aggr.	Campanula rotundifolia	Campanula rotundifolia	Landolt et al. (2010)	
Carex muricata aggr.	Carex divulsa	Carex divulsa	Landolt et al. (2010)	
Carex muricata aggr.	Carex muricata	Carex muricata	Landolt et al. (2010)	
Carex muricata aggr.	Carex muricata subsp. lamprocarpa	Carex muricata	Landolt et al. (2010)	
Carex muricata aggr.	Carex pairaei	Carex muricata subsp. pairae	Landolt et al. (2010)	
Carex muricata aggr.	Carex spicata	Carex spicata	Landolt et al. (2010)	
Centaurea ovina aggr.	Centaurea besserana	Centaurea besserana	Ad hoc	Centaurea ovina subsp. besseriana
Centaurea ovina aggr.	Centaurea lavrenkoana	Centaurea lavrenkoana	Ad hoc	Centaurea ovina subsp. lavrenkoana
Cotoneaster integerrimus aggr.	Cotoneaster integerrimus	Cotoneaster integerrimus	Landolt et al. (2010)	
Cotoneaster integerrimus aggr.	Cotoneaster matrensis	Cotoneaster melanocarpus	Landolt et al. (2010)	
Cotoneaster integerrimus aggr.	Cotoneaster melanocarpus	Cotoneaster melanocarpus	Landolt et al. (2010)	
Cotoneaster integerrimus aggr.	Cotoneaster tomentosus	Cotoneaster tomentosus	Landolt et al. (2010)	
Crambe tataria aggr.	Crambe aspera	Crambe aspera	Ad hoc	Crambe tataria var. buschii
Crambe tataria aggr.	Crambe pinnatifida	Crambe tataria	Ad hoc	Crambe tataria
Crambe tataria aggr.	Crambe tataria	Crambe tataria	Ad hoc	Crambe tataria
Cuscuta epithymum aggr.	Cuscuta approximata	Cuscuta approximata	Landolt et al. (2010)	
Cuscuta epithymum aggr.	Cuscuta epithymum	Cuscuta epithymum	Landolt et al. (2010)	
Cuscuta epithymum aggr.	Cuscuta epithymum subsp. kotschyi	Cuscuta epithymum	Landolt et al. (2010)	
Cytisus hirsutus aggr.	Chamaecytisus hirsutus	Cytisus hirsutus	Landolt et al. (2010)	
Cytisus hirsutus aggr.	Chamaecytisus polychrichus	Cytisus hirsutus subsp. polytrichus	Landolt et al. (2010)	
Cytisus hirsutus aggr.	Chamaecytisus ratisbonensis	Cytisus ratisbonensis	Landolt et al. (2010)	
Cytisus hirsutus aggr.	Chamaecytisus ruthenicus	Cytisus ruthenicus	Landolt et al. (2010)	
Cytisus hirsutus aggr.	Chamaecytisus supinus	Cytisus hirsutus	Landolt et al. (2010)	
Dianthus carthusianorum aggr.	Dianthus carpatius	Dianthus carthusianorum	Ehrendorfer (1973)	
Dianthus carthusianorum aggr.	Dianthus carthusianorum	Dianthus carthusianorum	Ehrendorfer (1973)	
Dianthus carthusianorum aggr.	Dianthus carthusianorum subsp. carthusianorum	Dianthus carthusianorum	Ehrendorfer (1973)	
Dianthus carthusianorum aggr.	Dianthus giganteiformis	Dianthus giganteiformis	Ehrendorfer (1973)	
Dianthus carthusianorum aggr.	Dianthus pontederae	Dianthus pontederae	Ehrendorfer (1973)	
Dianthus carthusianorum aggr.	Dianthus tenuifolius	Dianthus tenuifolius	Ehrendorfer (1973)	
Dianthus petraeus aggr.	Dianthus petraeus subsp. orbelicus	Dianthus petraeus subsp. orbelicus	Ad hoc	Dianthus petraeus subsp. orbelicus
Dianthus petraeus aggr.	Dianthus spiculifolius	Dianthus spiculifolius	Ad hoc	Dianthus petraeus subsp. spiculifolius
Dictamnus albus aggr.	Dictamnus albus	Dictamnus albus	Ad hoc	Dictamnus albus
Dictamnus albus aggr.	Dictamnus gymnostylis	Dictamnus gymnostylis	Ad hoc	Dictamnus albus subsp. gymnostylis
Euphrasia stricta aggr.	Euphrasia stricta	Euphrasia stricta	Ehrendorfer (1973)	
Euphrasia stricta aggr.	Euphrasia pectinata	Euphrasia pectinata	Ehrendorfer (1973)	
Festuca ovina aggr.	Festuca beckeri	Festuca beckeri	Ad hoc	Festuca ovina var. beckeri
Festuca ovina aggr.	Festuca callieri	Festuca callieri	Ad hoc	Festuca ovina var. callieri
Festuca ovina aggr.	Festuca dalmatica	Festuca dalmatica	Ehrendorfer (1973)	
Festuca ovina aggr.	Festuca ovina	Festuca ovina	Ehrendorfer (1973)	
Festuca valesiaca aggr.	Festuca pachyphylla	Festuca stricta subsp. rumelica	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca pseudodalmatica	Festuca pseudodalmatica	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca pseudovina	Festuca valesiaca subsp. parviflora	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca rupicola	Festuca stricta subsp. sulcata	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca rupicola subsp. rupicola	Festuca stricta subsp. sulcata	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca saxatilis	Festuca stricta subsp. saxatilis	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca stricta	Festuca stricta	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca sulcata	Festuca stricta subsp. sulcata	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca trachyphylla	Festuca stricta subsp. trachyphylla	Mosyakin & Fedoronchuk (1999)	
Festuca valesiaca aggr.	Festuca valesiaca	Festuca valesiaca	Mosyakin & Fedoronchuk (1999)	
Galium mollugo aggr.	Galium lucidum	Galium lucidum	Ehrendorfer (1973)	
Galium mollugo aggr.	Galium mollugo	Galium mollugo	Ehrendorfer (1973)	
Galium mollugo aggr.	Galium mollugo subsp. erectum	Galium mollugo subsp. erectum	Ehrendorfer (1973)	
Galium pusillum aggr.	Galium pumilum	Galium pumilum	Ehrendorfer (1973)	
Galium pusillum aggr.	Galium austriacum	Galium austriacum	Ehrendorfer (1973)	
Galium verum aggr.	Galium ruthenicum	Galium ruthenicum	Ad hoc	Galium verum subsp. ruthenicum
Galium verum aggr.	Galium verum	Galium verum	Ad hoc	Galium verum
Genista tinctoria aggr.	Genista tinctoria	Genista tinctoria	Ad hoc	Genista tinctoria
Genista tinctoria aggr.	Genista depressa	Genista depressa	Ad hoc	Genista tinctoria var. depressa
Genista tinctoria aggr.	Genista ovata	Genista tinctoria	Ad hoc	Genista tinctoria
Genista tinctoria aggr.	Genista tanaica	Genista tinctoria	Ad hoc	Genista tinctoria
Goniolimon tataricum aggr.	Goniolimon besseranum	Goniolimon besseranum	Ad hoc	Goniolimon tataricum subsp. besserianum
Goniolimon tataricum aggr.	Goniolimon tataricum	Goniolimon tataricum	Ad hoc	Goniolimon tataricum
Helianthemum canum aggr.	Helianthemum canum	Helianthemum canum	Ad hoc, based on Chusova & Didukh (2014)	Helianthemum canum
Helianthemum canum aggr.	Helianthemum canum subsp. canum	Helianthemum canum	Ad hoc, based on Chusova & Didukh (2014)	Helianthemum canum
Helianthemum canum aggr.	Helianthemum stevenii	Helianthemum stevenii	Ad hoc, based on Chusova & Didukh (2014)	Helianthemum canum subsp. stevenii
Helianthemum canum aggr.	Helianthemum creticola	Helianthemum creticola	Ad hoc, based on Chusova & Didukh (2014)	Helianthemum creticola
Helianthemum canum aggr.	Helianthemum cretophilum	Helianthemum cretophilum	Ad hoc, based on Chusova & Didukh (2014)	Helianthemum cretophilum
Hieracium lachenalii aggr.	Hieracium argillaceum	Hieracium lachenalii subsp. argillaceum	FloraVeg (2024)	
Hieracium lachenalii aggr.	Hieracium diaphanum	Hieracium diaphanum	FloraVeg (2024)	
Hierochloa odorata aggr.	Hierochloa australis	Hierochloa australis	Landolt et al. (2010)	
Hierochloa odorata aggr.	Hierochloa odorata	Hierochloa odorata	Landolt et al. (2010)	
Hierochloa odorata aggr.	Hierochloa repens	Hierochloa repens	Landolt et al. (2010)	
Inula salicina aggr.	Inula aspera	Inula aspera	Landolt et al. (2010)	
Inula salicina aggr.	Inula salicina	Inula salicina	Landolt et al. (2010)	
Inula salicina aggr.	Inula salicina subsp. salicina	Inula salicina	Landolt et al. (2010)	
Iris pumila aggr.	Iris pumila	Iris pumila	Ad hoc	Iris pumila
Iris pumila aggr.	Iris scariosa	Iris scariosa	Ad hoc	Iris pumila var. scariosa
Isatis tinctoria aggr.	Isatis praecox	Isatis praecox	Landolt et al. (2010)	
Isatis tinctoria aggr.	Isatis tinctoria	Isatis tinctoria	Landolt et al. (2010)	
Isatis tinctoria aggr.	Isatis tinctoria subsp. tinctoria	Isatis tinctoria	Landolt et al. (2010)	
Jurinea mollis aggr.	Jurinea arachnoidea	Jurinea arachnoidea	Ad hoc, based on Euro+Med (2024)	Jurinea mollis subsp. arachnoidea
Jurinea mollis aggr.	Jurinea calcarea	Jurinea calcarea	Ad hoc, based on Euro+Med (2024)	Jurinea calcarea
Jurinea mollis aggr.	Jurinea glycacantha	Jurinea glycacantha	Ad hoc, based on Euro+Med (2024)	Jurinea glycacantha
Jurinea mollis aggr.	Jurinea ledebourii	Jurinea ledebourii	Ad hoc, based on Euro+Med (2024)	Jurinea ledebourii
Jurinea mollis aggr.	Jurinea mollis	Jurinea mollis	Ad hoc, based on Euro+Med (2024)	Jurinea mollis
Jurinea mollis aggr.	Jurinea mollis subsp. mollis	Jurinea mollis	Ad hoc, based on Euro+Med (2024)	Jurinea mollis
Jurinea mollis aggr.	Jurinea mollis subsp. transylvanica	Jurinea mollis	Ad hoc, based on Euro+Med (2024)	Jurinea mollis
Jurinea mollis aggr.	Jurinea mollissima	Jurinea mollissima	Ad hoc, based on Euro+Med (2024)	Jurinea mollissima
Jurinea mollis aggr.	Jurinea sordida	Jurinea roegneri	Ad hoc, based on Euro+Med (2024)	Jurinea roegneri
Jurinea mollis aggr.	Jurinea talijevii	Jurinea cretacea	Ad hoc, based on Euro+Med (2024)	Jurinea cretacea
Jurinea mollis aggr.	Jurinea transylvanica	Jurinea transylvanica	Ad hoc, based on Euro+Med (2024)	Jurinea transylvanica
Koeleria brevis aggr.	Koeleria brevis	Koeleria brevis	Ad hoc	Koeleria lobata
Koeleria brevis aggr.	Koeleria lobata	Koeleria lobata	Ad hoc	Koeleria lobata
Koeleria macrantha aggr.	Koeleria cristata	Koeleria cristata	Ad hoc	Koeleria cristata
Koeleria macrantha aggr.	Koeleria gracilis	Koeleria macrantha	Ad hoc	Koeleria macrantha
Koeleria macrantha aggr.	Koeleria macrantha	Koeleria macrantha	Ad hoc	Koeleria macrantha
Leontodon crispus aggr.	Leontodon asper	Leontodon biscutellifolius	Euro+Med (2024)	
Leontodon crispus aggr.	Leontodon biscutellifolius	Leontodon biscutellifolius	Euro+Med (2024)	
Leontodon crispus aggr.	Leontodon crispus	Leontodon crispus	Euro+Med (2024)	
Leontodon crispus aggr.	Leontodon crispus subsp. crispus	Leontodon crispus	Euro+Med (2024)	
Linum flavum aggr.	Linum czerniaeii	Linum pallasianum	Ad hoc, based on Didukh et al. (2021)	Linum pallasianum
Linum flavum aggr.	Linum flavum	Linum flavum	Ad hoc, based on Mosyakin & Fedoronchuk (1999)	Linum flavum
Linum flavum aggr.	Linum ucranicum	Linum ucranicum	Ad hoc, based on Mosyakin & Fedoronchuk (1999)	Linum ucranicum
Luzula campestris aggr.	Luzula campestris	Luzula campestris	Landolt et al. (2010)	
Luzula campestris aggr.	Luzula multiflora	Luzula multiflora	Landolt et al. (2010)	
Melica ciliata aggr.	Melica ciliata	Melica ciliata	Ehrendorfer (1973)	
Melica ciliata aggr.	Melica ciliata subsp. ciliata	Melica ciliata	Ehrendorfer (1973)	
Melica ciliata aggr.	Melica transsilvanica	Melica transsilvanica	Ehrendorfer (1973)	
Myosotis arvensis aggr.	Myosotis arvensis	Myosotis arvensis	Ad hoc	Myosotis arvensis
Myosotis arvensis aggr.	Myosotis discolor	Myosotis discolor	Ad hoc	Myosotis arvensis var. versicolor
Odontarrhena tortuosa aggr.	Alyssum obtusifolium	Odontarrhena obtusifolia	Ad hoc	Alyssum tortuosum subsp. obtusifolium
Odontarrhena tortuosa aggr.	Alyssum tortuosum	Odontarrhena tortuosa	Ad hoc	Odontarrhena tortuosa
Onobrychis vicifolia aggr.	Onobrychis arenaria	Onobrychis arenaria	Ehrendorfer (1973)	
Onobrychis vicifolia aggr.	Onobrychis miniata	Onobrychis arenaria subsp. miniata	Ehrendorfer (1973)	
Onobrychis vicifolia aggr.	Onobrychis tanaitica	Onobrychis arenaria	Ehrendorfer (1973)	
Onobrychis vicifolia aggr.	Onobrychis vicifolia	Onobrychis vicifolia	Ehrendorfer (1973)	
Onosma visianii aggr.	Onosma macrochaeta	Onosma visianii	Ad hoc	Onosma visianii
Onosma visianii aggr.	Onosma pseudarenaria	Onosma pseudorenaria	Ad hoc	Onosma visianii subsp. transsilvanica
Onosma visianii aggr.	Onosma taurica	Onosma pseudorenaria	Ad hoc	Onosma visianii subsp. transsilvanica
Onosma visianii aggr.	Onosma visianii	Onosma visianii	Ad hoc	Onosma visianii
Pilosella piloselloides aggr.	Hieracium bauhini	Pilosella bauhini	Euro+Med (2024)	
Pilosella piloselloides aggr.	Hieracium cymanthum	Pilosella bauhini	Euro+Med (2024)	
Pilosella piloselloides aggr.	Hieracium pavichii	Pilosella pavichii	Euro+Med (2024)	
Pilosella piloselloides aggr.	Hieracium piloselloides	Hieracium piloselloides	Euro+Med (2024)	
Pilosella piloselloides aggr.	Hieracium praetulum subsp. bauhini	Pilosella bauhini	Euro+Med (2024)	
Pimpinella saxifraga aggr.	Pimpinella major	Pimpinella major	Juillerat et al. (2017)	
Pimpinella saxifraga aggr.	Pimpinella nigra	Pimpinella nigra	Juillerat et al. (2017)	
Pimpinella saxifraga aggr.	Pimpinella saxifraga	Pimpinella saxifraga	Juillerat et al. (2017)	
Potentilla argentea aggr.	Potentilla argentea	Potentilla argentea	Ad hoc	Potentilla argentea
Potentilla argentea aggr.	Potentilla inclinata	Potentilla inclinata	Ad hoc	Potentilla argentea subsp. inclinata
Potentilla incana aggr.	Potentilla arenaria	Potentilla incana	Ad hoc, based on Didukh et al. (2021)	Potentilla incana
Potentilla incana aggr.	Potentilla incana	Potentilla incana	Ad hoc, based on Didukh et al. (2021)	Potentilla incana
Potentilla incana aggr.	Potentilla tabernaemontani	Potentilla tabernaemontani	Ad hoc, based on Didukh et al. (2021)	Potentilla tabernaemontani
Prunella vulgaris aggr.	Potentilla tommasiniana	Potentilla tommasiniana	Ad hoc, based on Didukh et al. (2021)	Potentilla incana subsp. tommasiniana
Prunella vulgaris aggr.	Prunella laciniata	Prunella laciniata	Ad hoc	Prunella vulgaris var. laciniata
Prunella vulgaris aggr.	Prunella vulgaris	Prunella vulgaris	Ad hoc	Prunella vulgaris
Psephellus marschallianus aggr.	Centaurea carbonata	Psephellus carbonatus	Ad hoc, based on Didukh et al. (2018)	Psephellus carbonatus
Psephellus marschallianus aggr.	Centaurea marschalliana	Psephellus marschallianus	Ad hoc, based on Didukh et al. (2018)	Psephellus marschallianus
Psephellus marschallianus aggr.	Centaurea sumensis	Psephellus sumensis	Ad hoc, based on Didukh et al. (2018)	Psephellus sumensis
Rosa canina aggr.	Rosa canina	Rosa canina	Landolt et al. (2010)	
Rosa canina aggr.	Rosa corymbifera	Rosa corymbifera	Landolt et al. (2010)	
Salvia pratensis aggr.	Salvia pratensis	Salvia pratensis	Ad hoc	Salvia pratensis
Salvia pratensis aggr.	Salvia stepposa	Salvia dumetorum	Ad hoc	Salvia pratensis var. dumetorum
Satureja montana aggr.	Satureja montana subsp. kitaibelii	Satureja kitaibelii	Ad hoc	Satureja montana subsp. kitaibelii
Satureja montana aggr.	Satureja taurica	Satureja taurica	Ad hoc	Satureja montana subsp. taurica
Senecio squalidus aggr.	Senecio rupestris	Senecio rupestris	Ad hoc	Senecio rupestris
Senecio squalidus aggr.	Senecio squalidus	Senecio squalidus	Ad hoc	Senecio squalidus
Sesleria rigida aggr.	Sesleria filifolia	Sesleria filifolia	Ad hoc	Sesleria rigida var. panicii
Sesleria rigida aggr.	Sesleria rigida	Sesleria rigida	Ad hoc	Sesleria rigida
Sesleria rigida aggr.	Sesleria rigida subsp. rigida	Sesleria rigida	Ad hoc	Sesleria rigida
Silene italica aggr.	Silene italica	Silene italica	Ehrendorfer (1973)	
Silene italica aggr.	Silene italica subsp. italica	Silene italica	Ehrendorfer (1973)	
Silene italica aggr.	Silene italica subsp. nemoralis	Silene italica	Ehrendorfer (1973)	
Silene italica aggr.	Silene nemoralis	Silene nemoralis	Ehrendorfer (1973)	
Silene otites aggr.	Otites borysthonica	Silene borysthonica	Ehrendorfer (1973)	
Silene otites aggr.	Otites eugeniae	Silene eugeniae	Ad hoc	Silene eugeniae
Silene otites aggr.	Sil			

Supplement E2. Life forms, Grime’s strategy for the species list used in the analysis.

Anhang E2. Lebensformen, Grime’s Strategie für die Artenliste, die in der Analyse genutzt wurden.

Species name	Life formes	Grime' strategy	Species name	Life formes	Grime' strategy
Acanthus hungaricus	hem	C	Clinopodium suaveolens	hem	S
Acer campestre	pha	C	Clinopodium vulgare	hem	C
Acer negundo	pha	CR	Consolida regalis	the	R
Acer platanoides	pha	C	Convallaria majalis	geo	C
Acer pseudoplatanus	pha	C	Convolvulus arvensis	geo	CR
Acer tataricum	pha	C	Convolvulus calvertii subsp. calverti	cha	S
Achillea coarctata	hem	C	Convolvulus cantabrica	cha	CS
Achillea crithmifolia	hem	C	Convolvulus fruticosus	cha	CS
Achillea millefolium aggr.	hem	C	Convolvulus lineatus	cha	CS
Achillea nobilis	hem	C	Convolvulus sericecephalus	cha	S
Achnatherum splendens	hem	CS	Cornus mas	pha	C
Aconitum anthora	hem	C	Cornus sanguinea	pha	C
Aconitum moldavicum	hem	C	Coronilla coronata	hem	CS
Adonis vernalis	hem	C	Coronilla scorpioides	the	CRS
Adonis volgensis	hem	C	Coronilla vaginalis	cha	CS
Aegilops triuncialis	the	R	Corydalis cava	geo	C
Agrimonia eupatoria aggr.	hem	C	Corydalis solida	geo	C
Agrimonia repens	hem	C	Corylus avellana	pha	C
Agropyron cristatum aggr.	hem	CS	Cota tinctoria	hem	CS
Agrostis capillaris	hem	CS	Cotinus coggygia	pha	CS
Ailanthus altissima	pha	CRS	Cotoneaster integerrimus aggr.	pha	CS
Ajuga chamaeptytis	the	RS	Cotoneaster sp.	pha	
Ajuga genevensis	hem	C	Cousinia astracanica	cha	S
Ajuga laxmannii	hem	C	Crambe tataria aggr.	cha	CS
Alchemilla jailae	hem	C	Crataegus laevigata	pha	C
Alliaria petiolata	hem	CR	Crataegus monogyna	pha	C
Allium ascalonicum	geo	R	Crataegus rhipidophylla	pha	C
Allium atroviolaceum	geo	CS	Crepis foetida	the	R
Allium delicatulum	geo	S	Crepis jacquinii	hem	C
Allium denudatum	geo	S	Crepis neglecta	the	RS
Allium ericetorum	geo	S	Cruciata glabra	hem	CS
Allium flavum	geo	CS	Cruciata taurica	cha	CS
Allium inaequale	geo	CS	Crupina vulgaris	the	RS
Allium inderiense	geo	CS	Cuscuta campestris	the	R
Allium lineare	geo	CS	Cuscuta epithymum aggr.	the	RCS
Allium lusitanicum aggr.	geo	CS	Cuscuta europaea	the	CR
Allium marschalianum	geo	CS	Cuscuta sp.	the	
Allium moschatum	geo	S	Cyanus pinnatifidus	hem	S
Allium obliquum	geo	S	Cyanus triumfettii	hem	CS
Allium oleraceum	geo	RS	Cynodon dactylon	geo	CRS
Allium paniculatum aggr.	geo	S	Cynoglossis barrelieri	hem	CS
Allium podolicum	geo	S	Cynosurus cristatus	hem	C
Allium rotundum	geo	CS	Cystopteris fragilis	hem	CS
Allium saxatile	geo	S	Cytisus albus	pha	CS
Allium schoenoprasum	geo	CS	Cytisus austriacus	pha	CS
Allium sp.	geo	CS	Cytisus blockianus	pha	CS
Allium sphaerocephalon	geo	CS	Cytisus graniticus	pha	CS
Allium tulipifolium	geo	CS	Cytisus hirsutus aggr.	cha	CS
Alopecurus vaginatus	hem	CS	Cytisus paczoskii	pha	C
Alyssum alyssoides	the	RS	Cytisus podolicus	pha	CS
Alyssum lenense	cha	CS	Cytisus procumbens	cha	CS
Alyssum montanum aggr.	hem	CS	Cytisus sp.	cha	
Alyssum turkestanicum	the	RS	Dactylis glomerata	hem	C
Amaranthus retroflexus	the	R	Dactylorhiza sambucina	geo	C
Amelanchier ovalis	pha	R	Danthonia alpina	hem	CS
Anabasis cretacea	cha	S	Daphne cneorum	cha	CS
Anabasis salsa	cha	SRS	Daucus carota	hem	CR
Anacamptis morio	geo	CS	Deschampsia cespitosa	hem	C
Anacamptis pyramidalis	geo	C	Descurainia sophia	the	R
Anagallis arvensis	the	R	Dianthus carthusianorum aggr.	hem	C
Androsace elongata	the	CR	Dianthus giganteus	hem	CS
Androsace maxima	the	CR	Dianthus lanceolatus	hem	CS
Androsace septentrionalis	the	RS	Dianthus henteri	hem	C
Androsace villosa	cha	CS	Dianthus nitidus	hem	C
Anemone nemorosa	geo	C	Dianthus petraeus aggr.	hem	CR
Anemone ranunculoides	geo	C	Dianthus plumarius	hem	CS
Anemone sylvestris	hem	C	Dianthus praecox	hem	CS
Anisantha rigida	the	RCS	Dianthus pseudarmeria	the	RS
Anisantha tectorum	the	R	Dianthus ramosissimus	cha	CS
Antennaria dioica	cha	CS	Dichoropetalum carvifolia	hem	C
Anthemis arvensis	the	CR	Dictamnus albus aggr.	hem	CS
Anthericum ramosum	geo	CS	Digitalis grandiflora	hem	C
Anthoxanthum odoratum	hem	CR	Diplotaxis cretacea	hem	S
Anthyllis vulneraria aggr.	hem	CS	Diplotaxis muralis	the	CR
Aquilegia subcaposa	hem	C	Diplotaxis tenuifolia	hem	CR
Aquilegia vulgaris	hem	C	Doronium columnae	hem	CS
Arabidopsis arenosa	hem	RS	Dorycnium pentaphyllum	cha	CS
Arabidopsis petraea	hem	S	Draba aizoides	hem	S
Arabidopsis thaliana	the	RS	Draba cuspidata	cha	S
Arabis auriculata	the	CR	Draba lasiocarpa	hem	S
Arabis hirsuta	hem	CR	Draba nemorosa	the	CRS
Arabis procurrens	cha	S	Draba podolica	hem	S
Arabis sagittata	hem	CR	Draba verna	the	RS
Arabis sp.	hem		Dracocephalum austriacum	hem	CS
Archanthemis trotzkiana	cha	S	Dryomacallis geoides	hem	S
Arenaria koriniana	hem	CS	Echinops bannaticus	hem	CS
Arenaria procrea	hem	CS	Echinops ritro	hem	CS
Arenaria rigida	hem	CS	Echinops sphaerocephalus	hem	CS
Arenaria serpyllifolia aggr.	the	RS	Echium vulgare	hem	CRS
Aristolochia clematitis	geo	CS	Edraianthus graminifolius	hem	CS
Aristolochia pallida	geo	CS	Elymus caninus	hem	C
Arrhenatherum elatius	hem	CR	Elytrigia caespitosa subsp. nodosa	hem	CS
Artemisia absinthium	hem	CRS	Elytrigia intermedia	geo	C
Artemisia alba	cha	CS	Elytrigia repens	geo	C
Artemisia alpina	cha	S	Elytrigia stipifolia	hem	C
Artemisia austriaca	cha	CS	Elytrigia strigosa	hem	CS
Artemisia campestris	hem	CS	Ephedra distachya	cha	CS
Artemisia dracunculus	hem	C	Epipactis atrorubens	geo	C
Artemisia hololeuca	cha	S	Eremopyrum triticeum	the	CRS
Artemisia lerciana	cha	CS	Eremurus spectabilis	hem	CS
Artemisia lessingiana	cha	S	Erigeron acris	hem	CR
Artemisia nitrosa	cha	CS	Erigeron canadensis	the	CR
Artemisia nutans	cha	S	Eriosynaphe longifolia	cha	CS
Artemisia sansloides	cha	S	Erodium cicutarium	the	RS
Artemisia santonicum	cha	CS	Erucastrum nasturtiifolium	hem	R
Artemisia semiarida	cha	CS	Eryngium campestre	hem	C
Artemisia taurica	cha	CS	Eryngium planum	hem	C
Artemisia terrae-albae	cha	CS	Erysimum canum	hem	CRS
Asarum europaeum	geo	C	Erysimum comatum	hem	CS
Asparagus iherensis	hem	CS	Erysimum cretaceum	hem	S
Asparagus officinalis	geo	C	Erysimum cuspidatum	hem	CRS
Asparagus tenuifolius	geo	C	Erysimum diffusum	hem	R
Asperula arvensis	the	S	Erysimum leptostylum	hem	CS
Asperula capitata	hem	S	Erysimum odoratum	hem	CS
Asperula cretacea	cha	S	Erysimum rhaeticum	hem	RS
Asperula cynanchica	hem	CS	Erysimum sp.	hem	
Asperula montana	cha	CS	Erysimum talijevii	hem	S
Asperula purpurea	hem	S	Erysimum ucrainicum	hem	S
Asperula supina	cha	S	Erysimum witmannii	hem	S
Asperula taurica	hem	CS	Euonymus verrucosus	pha	C
Asperula tenella	hem	CS	Euphorbia agraria	hem	CR
Asperula tephrocarpa	cha	S	Euphorbia amygdaloides	hem	C
Asperula tinctoria	hem	C	Euphorbia angulata	hem	C
Asphodeline lutea	geo	CS	Euphorbia cyparissias	hem	C
Asphodeline taurica	geo	CS	Euphorbia epithymoides	hem	C
Asplenium ceterach	hem	S	Euphorbia esula	hem	CR
Asplenium ruta-muraria	hem	S	Euphorbia myrsinites	cha	S
Asplenium septentrionale	hem	S	Euphorbia nicaeensis	hem	S
Asplenium trichomanes	hem	S	Euphorbia petrophila	hem	S
Asplenium viride	hem	S	Euphorbia seguieriana	hem	CS
Aster alpinus	hem	CS	Euphorbia semivillosa	hem	C
Aster amellus	hem	C	Euphorbia stricta	the	CR
Astragalus aktubensis	cha	S	Euphrasia hirtella	the	CR
Astragalus albicaulis	cha	CS	Euphrasia rostkoviana	the	CR
Astragalus austriacus	hem	CS	Euphrasia salisburgensis	the	CRS
Astragalus brachylobus	cha	CS	Euphrasia stricta aggr.	the	CR
Astragalus buchtormensis	hem	CS	Euphrasia taurica	the	CRS
Astragalus cornutus	cha	CS	Fagus sylvatica	pha	C
Astragalus danicus	hem	C	Falcaria vulgaris	hem	CS
Astragalus glaucus	cha	CS	Fallopia convolvulus	the	RS
Astragalus glycyphyllos	hem	C	Fallopia dumetorum	the	RS
Astragalus kystanaicus	cha	S	Ferula caspica	hem	S
Astragalus leontinus	hem	CS	Ferula tatarica	hem	CS
Astragalus medius	cha	CS	Ferulago campestris	hem	C
Astragalus monspesulanus	hem	CS	Ferulago sylvatica	hem	C
Astragalus onobrychis	hem	CS	Ferulago taurica	hem	CS
Astragalus ponticus	hem	CS	Festuca cretacea	hem	CS
Astragalus pseudoglaucus	cha	CS	Festuca glauca	hem	CS
Astragalus roemerii	hem	CS	Festuca heterophylla	hem	CS
Astragalus rupifragus	cha	S	Festuca ovina aggr.	hem	C
Astragalus stenoceras	cha	S	Festuca pallens	hem	CS
Astragalus storozevae	cha	CS	Festuca rubra	hem	CS
Astragalus subarcuatus	cha	S	Festuca valesiaca aggr.	hem	C
Astragalus subuliformis	cha	CS	Filago arvensis	the	RS
Astragalus sulcatus	hem	CS	Filago germanica	the	RS
Astragalus temirensis	cha	CS	Filipendula vulgaris	hem	C
Astragalus tenuifolius	cha	CS	Fragaria moschata	hem	C
Astragalus testiculatus	hem	CS	Fragaria vesca	hem	C
Astragalus varius	cha	CS	Fragaria viridis	hem	C
Astragalus vesicarius aggr.	cha	CS	Frangula alnus	pha	C
Asyneuma canescens	hem	C	Fraxinus excelsior	pha	C
Atocion armeria	the	RS	Fraxinus lanceolata	pha	
Atraphaxis decipiens	cha	S	Fraxinus ornus	pha	C
Atraphaxis frutescens	cha	S	Fritillaria orientalis	geo	S
Atraphaxis replicata	cha	S	Fumana procumbens	cha	CS
Atraphaxis spinosa	cha	S	Fumaria officinalis	the	R
Aurinia petraea	hem	S	Gagea bulbifera	geo	CS
Aurinia saxatilis	hem	S	Gagea pratensis	geo	C
Avenula pubescens	hem	C	Gagea pusilla	geo	C
Barbarea vulgaris	hem	RS	Galanthus nivalis	geo	C
Bassia prostrata	cha	CS	Galatella angustissima	hem	CS
Bellidiastrum michelii	hem	CS	Galatella linoxyris	hem	C
Berberis vulgaris	pha	CR	Galatella tatarica	hem	CS
Berteroia incana	the	CRS	Galatella villosa	hem	CS
Betula pendula	pha	CRS	Galeopsis angustifolia	the	R
Biscutella laevigata	hem	CS	Galeopsis ladanum	the	R
Bothriochloa ischaemum	hem	CS	Galitzkya spathulata	hem	S
Botrychium lunaria	geo	C	Galium album	hem	CS
Brachypodium pinnatum	hem	C	Galium anisophyllum	hem	RS
Brassica elongata	hem	S	Galium aparine	the	CR
Briza media	hem	C	Galium baillonii	hem	CS
Bromopsis benekenii	hem	CS	Galium biebersteinii	hem	CS
Bromopsis cappadocica	hem	CS	Galium boreale	hem	C
Bromopsis erecta	hem	CS	Galium cracoviense	hem	S
Bromopsis inermis	hem	C	Galium glaucum	hem	CS
Bromopsis pannonica	hem	C	Galium humifusum	hem	RS
Bromopsis ramosa	hem	C	Galium mollugo aggr.	hem	C
Bromopsis riparia	hem	CS	Galium octonarium	hem	CS
Bromus arvensis	the	R	Galium pumilum aggr.	hem	C
Bromus commutatus	the	R	Galium transcarpaticum	hem	CS
Bromus hordeaceus	the	R	Galium verum aggr.	hem	CS
Bromus japonicus	the	RS	Galium volhyanicum	hem	CS
Bromus squarrosus	the	R	Genista alba	cha	CS
Buglossoides arvensis	the	R	Genista januensis	cha	S
Bupleurum falcatum	hem	CS	Genista pilosa	cha	CS
Bupleurum longifolium	hem	CS	Genista sagittalis	cha	C
Bupleurum pachnospermum	the	RS	Genista scythica	cha	CS
Calamagrostis epiglouca	hem	C	Genista tetragona	cha	S
Calamagrostis eripiojeas	hem	CS	Genista tinctoria aggr.	cha	CS
Calamagrostis sp.	hem		Gentiana clusii	hem	CS
Calamagrostis varia	hem	C	Gentiana cruciata	hem	C
Camelina microcarpa	the	CRS	Gentianella lutescens	hem	CRS
Campanula bononiensis	hem	C	Geranium columbinum	the	R
Campanula carpatica	hem	CS	Geranium lucidum	the	CRS
Campanula crassipes	hem	S	Geranium pusillum	the	R
Campanula glomerata	hem	C	Geranium robertianum	the	CRS
Campanula lingulata	hem	CS	Geranium rotundifolium	the	R
Campanula patula	hem	C	Geranium sanguineum	hem	C
Campanula persicifolia	hem	C	Geranium sp.	hem	
Campanula rapunculoides	hem	C	Geranium sylvaticum	hem	C
Campanula rotundifolia aggr.	hem	CS	Geum urbanum	hem	CR
Campanula sibirica	hem	CS	Glechoma hederacea	hem	CR
Campanula trachelium	hem	C	Glechoma hederacea	hem	C
Camphorosma monspeliaca	cha	R	Globularia bisnagarica	hem	CS
Capsella bursa-pastoris	the	CR	Goniolimon rubellum	hem	CS
Caragana balchashensis	npha	CS	Goniolimon speciosum	hem	CS
Caragana frutex	npha	C	Goniolimon tataricum aggr.	hem	CS
Carduus acanthoides	the	R	Gymnadenia conopsea	geo	CS
Carduus candicans	hem	CR	Gymnadenia odoratissima	geo	CS
Carduus collinus	hem	CS	Gymnocarpium robertianum	geo	CS
Carduus defloratus	hem	CS	Gypsophila altissima	hem	CS
Carduus nutans	hem	CR	Gypsophila collina	cha	CS
Carex alba	hem	CS	Gypsophila diffusa	cha	S
Carex brevicollis	hem	C	Gypsophila fastigiata	hem	CR
Carex caryophylla	hem	C	Gypsophila glomerata	hem	S
Carex cespitosa	hem	C	Gypsophila muralis	the	CS
Carex digitata	hem	C	Gypsophila oligosperma	hem	CS
Carex filiformis	hem	C	Gypsophila pallasi	hem	S
Carex flacca	hem	C	Gypsophila paniculata	hem	CRS
Carex halleriana	hem	C	Gypsophila petraea	cha	S
Carex humilis	hem	CS	Gypsophila repens	cha	S
Carex liparocarpos	hem	CS	Gypsophila rupestris	cha	S
Carex michelii	hem	C	Gypsophila thyraica	cha	CS
Carex montana	hem	CS	Haplophyllum suaveolens	hem	CS
Carex muricata aggr.	hem	C	Hedysarum argyrophyllum	cha	CS
Carex ornithopoda	hem	CS	Hedysarum candidum	hem	CS
Carex pediformis	hem	CS	Hedysarum cretaceum	hem	CS
Carex praecox	hem	CRS	Hedysarum grandiflorum	hem	CS
Carex supina	hem	CS	Hedysarum polychromum	cha	CS
Carlina acanthifolia subsp. utzka	hem	CS	Hedysarum razoumovianum	cha	CS
Carlina acaulis	hem	CS	Hedysarum tscherkassovae	cha	S
Carlina biebersteinii	hem	R	Hedysarum ucrainicum	hem	S
Carlina sp.	hem		Helianthemum alpestre	cha	CS
Carlina vulgaris	hem	R	Helianthemum canum aggr.	cha	CS
Centaurea atrorupurea	hem	CS	Helianthemum nummularium	cha	CS
Centaurea diffusa	hem	R	Helianthemum orientale	cha	CS
Centaurea jacea	hem	C	Helianthemum salicifolium	the	CR
Centaurea orientalis	hem	CS	Helianthemum sp.	cha	
Centaurea ovina aggr.	hem	S	Helichrysum arenarium	hem	CS
Centaurea reichenbachii	hem	CS	Helichrysum graveolens	hem	

Species name	Life formes	Grime' strategy	Species name	Life formes	Grime' strategy
Inula britannica	hem	C	Psephellus trinervius	hem	CS
Inula conyzae	hem	C	Pseudopodospermum hispanicum	hem	CS
Inula ensifolia	hem	CS	Pseudosedum lievenii	cha	S
Inula hirta	hem	C	Pseudoturnitis turrita	hem	C
Inula oculus-christi	hem	C	Puccinellia distans	hem	CS
Inula salicina aggr.	hem	C	Pulsatilla halleri	hem	CS
Inula spiraeifolia	hem	C	Pulsatilla montana	hem	CS
Ionopsidium abulense	the	S	Pulsatilla patens	hem	C
Iris aphylla	hem	CS	Pulsatilla pratensis	hem	C
Iris pontica	geo	C	Pulsatilla sp.	hem	C
Iris pumila aggr.	hem	CS	Pulsatilla vulgaris	hem	C
Iris reichenbachii	geo	CS	Pyrus communis	pha	CR
Iris ruthenica	hem	CS	Quercus cerris	pha	CS
Iris variegata	geo	CR	Quercus petraea	pha	CS
Isatis costata	hem	RS	Quercus pubescens	pha	CS
Isatis gymnocarpa	the	R	Ranunculus breyninus	hem	C
Isatis tinctoria aggr.	hem	CS	Ranunculus bulbosus	geo	C
Ixiolirion tataricum	geo	S	Ranunculus illyricus	geo	C
Jacobaea erucifolia	hem	CS	Ranunculus montanus	hem	C
Jacobaea vulgaris	hem	RS	Ranunculus polyanthemus	hem	C
Jovibarba globifera	cha	S	Rapistrum perenne	hem	RS
Jovibarba heuffelii	cha	S	Reseda lutea	hem	RS
Juglans regia	pha	CR	Reseda phyteuma	the	RS
Juniperus communis	pha	CS	Rhammatophyllum pachyrhizum	the	RS
Juniperus sabina	pha	CS	Rhamnus cathartica	pha	CRS
Juniperus sp.	pha	C	Rhamnus saxatilis subsp. tinctoria	pha	CS
Jurinea kirghisorum	cha	S	Rhamnus sp.	pha	C
Jurinea mollis aggr.	hem	CS	Rhaponticoides kasakorum	cha	S
Jurinea mugodsharica	cha	S	Rhaponticoides ruthenica	hem	CS
Jurinea multiflora	hem	CS	Rhaponticoides talievii	hem	CS
Jurinea stoechadifolia	cha	CS	Rhinanthus angustifolius	the	CR
Kernera saxatilis	hem	S	Rhinanthus minor	the	R
Klasea cardunculus	hem	CS	Rhinanthus rumelicus	the	R
Klasea erucifolia	hem	CS	Rindera tetraspis	hem	CR
Klasea radiata	hem	C	Rochelia disperma subsp. retorta	the	RS
Knautia arvensis	hem	C	Rosa canina aggr.	pha	CR
Knautia kitaibelii	hem	S	Rosa gallica	pha	C
Koeleria brevis aggr.	hem	CS	Rosa micrantha	pha	CS
Koeleria glauca	hem	C	Rosa mollis	pha	C
Koeleria grandis	hem	C	Rosa pendulina	pha	C
Koeleria macrantha aggr.	hem	C	Rosa rubiginosa	pha	C
Koeleria pyramidata	hem	C	Rosa sp.	pha	C
Koeleria sclerophylla	hem	C	Rosa spinosissima	pha	CS
Koeleria talievii	hem	CS	Rosa tomentosa	pha	C
Krascheninnikovia ceratoides	cha	S	Rubus canescens	pha	CR
Lactuca perennis	hem	CR	Rubus silesiacus	pha	CR
Lactuca quercina	hem	CR	Rumex acetosa	hem	C
Lactuca serriola	hem	CR	Rumex acetosella	hem	CRS
Lactuca viminea	hem	CR	Salix caprea	pha	C
Lamium alexicaule	the	R	Salsola tamariscina	the	RS
Lamium galeobdolon	cha	C	Salsola tragus	the	RS
Lamium maculatum	hem	C	Salvia nemorosa	hem	C
Lamium purpureum	the	CR	Salvia nutans	hem	CS
Lappula barbata	the	R	Salvia pratensis aggr.	hem	C
Lappula microcarpa	the	R	Salvia scabiosifolia	cha	CS
Lappula patula	the	R	Salvia verbenaca	hem	C
Lappula squarrosa	the	R	Salvia verticillata	hem	CRS
Laserpitium krapfii	hem	S	Sanguisorba minor	hem	CS
Laserpitium latifolium	hem	C	Sanguisorba officinalis	hem	C
Lathyrus nissolia	the	C	Saponaria bellidifolia	hem	CS
Lathyrus pannonicus	hem	C	Saponaria glutinosa	the	CS
Lathyrus pratensis	hem	C	Satureja montana aggr.	cha	S
Lathyrus sphaericus	the	R	Saxifraga adscendens	hem	S
Lathyrus vernus	geo	C	Saxifraga marginata	cha	S
Lembotropis nigricans	pha	CS	Saxifraga paniculata	hem	S
Leontodon crispus aggr.	hem	CS	Saxifraga tridactylites	the	RS
Leontodon hispidus	hem	CS	Scabiosa canescens	hem	CS
Leontodon incanus	hem	CS	Scabiosa columbaria	hem	C
Leontopodium nivale subsp. alpinum	hem	S	Scabiosa lucida	hem	CS
Leonurus quinquelobatus	hem	R	Scabiosa ochroleuca	hem	C
Lepidium campestre	hem	CR	Schedonorus arundinaceus	hem	CS
Lepidium coronopifolium	hem	CR	Schedonorus pratensis	hem	C
Lepidium draba	geo	CR	Scleranthus annuus	the	CRS
Lepidium meyeri	cha	S	Scleranthus perennis	hem	CS
Lepidium perfoliatum	the	RS	Scorzonera laciniata	hem	CRS
Leucanthemum vulgare	hem	C	Scorzonera mollis	hem	C
Leymus akmolinsensis	hem	S	Scorzonera purpurea	hem	C
Ligularia carpathica	hem	C	Scorzoneroides autumnalis	hem	C
Ligustrum vulgare	pha	C	Scrophularia cretacea	cha	S
Lilium martagon	geo	C	Scrophularia heterophylla subsp. lac	hem	S
Limonium cretaceum	cha	S	Scrophularia rupestris	cha	S
Limonium meyeri	cha	CS	Scutellaria albidia	hem	S
Limonium sp.	hem	C	Scutellaria orientalis	cha	S
Limonium suffruticosum	cha	CS	Scutellaria supina	cha	S
Linaria angustissima	hem	RS	Securigera elegans	hem	CRS
Linaria cretacea	geo	S	Securigera varia	hem	C
Linaria genistifolia	hem	S	Sedum acre	cha	CS
Linaria incompleta	hem	S	Sedum album	cha	CS
Linaria macroura	hem	CRS	Sedum annuum	the	RS
Linaria vulgaris	hem	CRS	Sedum hispanicum	the	S
Linum austriacum	hem	CS	Sedum rubens	the	S
Linum catharticum	the	CR	Sedum rupestre	cha	S
Linum flavum aggr.	cha	CS	Sedum sexangulare	cha	CS
Linum hirsutum	hem	CS	Selaginella helvetica	cha	CS
Linum nervosum	hem	CS	Selinum silaifolium	hem	CS
Linum perenne	hem	CRS	Sempervivum marmoreum	cha	S
Linum tauricum	cha	CS	Sempervivum ruthenicum	cha	CS
Linum tenuifolium	hem	CRS	Sempervivum tectorum	cha	S
Linum trigynum	the	CRS	Senecio glaucus subsp. coronopifoli	the	RS
Linum ucranicum	cha	CS	Senecio leucanthemifolius subsp. ve	the	R
Lithospermum officinale	hem	CR	Senecio sp.	hem	C
Litwinowia tenuissima	the	RS	Senecio squalidus aggr.	hem	RS
Lolium perenne	hem	CR	Senecio umbrosus	hem	C
Lomelosia argentea	hem	CRS	Seseli annuum	hem	CR
Lomelosia isetensis	cha	S	Seseli campestre	hem	C
Lomelosia micrantha	the	RS	Seseli dichotomum	hem	S
Lomelosia rotata	the	RS	Seseli eriocephalum	hem	S
Lotus corniculatus	hem	C	Seseli glabratum	cha	S
Luzula campestris aggr.	hem	C	Seseli gracile	hem	CS
Malva thuringiaca	hem	CR	Seseli gummiferum	cha	S
Marrubium peregrinum	hem	CRS	Seseli hippomarathrum	hem	S
Marrubium vulgare	hem	RS	Seseli lehmannii	hem	S
Matthiola fragrans	cha	S	Seseli leucospermum	hem	S
Matthiola odoratissima	cha	S	Seseli libanotis	hem	CS
Matthiola tatarica	cha	S	Seseli osseum	hem	CS
Medicago falcata	hem	CS	Seseli pallasii	hem	CS
Medicago lupulina	the	CR	Seseli rigidum	hem	S
Medicago minima	the	CR	Seseli sp.	hem	C
Medicago monspeliaca	the	CS	Seseli tortuosum	hem	CS
Medicago prostrata	hem	C	Sesleria caerulea	hem	CS
Medicago rupestris	cha	S	Sesleria heufflerana	hem	CS
Medicago sativa	hem	C	Sesleria rigida aggr.	hem	C
Melampyrum arvense	the	R	Setaria pumila	the	R
Melampyrum barbatum	the	R	Setaria viridis	the	R
Melampyrum bihariense	hem	C	Sherardia arvensis	the	RS
Melampyrum cristatum	the	C	Sideritis montana	the	CS
Melampyrum nemorosum	the	C	Sideritis taurica	cha	CS
Melica ciliata aggr.	hem	CS	Silene antri-jovis	cha	S
Melilotus albus	hem	RS	Silene bupleuroides	hem	CS
Melilotus officinalis	hem	R	Silene chlorantha	hem	C
Melilotus sp.	hem	C	Silene cretacea	cha	S
Melilotus sulcatus	the	R	Silene csereii	hem	RS
Meniocus linifolius	the	RS	Silene flaves cens	hem	S
Mercurialis perennis aggr.	hem	C	Silene italica aggr.	hem	CS
Minuartia glomerata	the	CRS	Silene latifolia	hem	CRS
Minuartia hirsuta	cha	CS	Silene nutans	hem	C
Minuartia rubra	the	RS	Silene otites aggr.	hem	CRS
Minuartia setacea	cha	S	Silene procumbens	hem	CS
Minuartia sp.	cha	C	Silene sp.	hem	C
Minuartia verna	cha	S	Silene supina	cha	CS
Misopates orontium	the	R	Silene viridiflora	hem	C
Moehringia muscosa	hem	CS	Silene viscaria	hem	C
Muscari comosum	geo	CS	Silene viscosa	the	CS
Muscari neglectum	geo	CS	Silene vulgaris	hem	CR
Muscari tenuiflorum	geo	CS	Silene zawadzki	hem	S
Myosotis alpestris	hem	C	Sisymbrium loeselii	the	R
Myosotis arvensis aggr.	hem	R	Sisymbrium polymorphum	the	CS
Myosotis lithospermifolia	hem	C	Solanum nigrum	the	R
Myosotis ramosissima	the	CRS	Solidago virgaurea	hem	CS
Myosotis sparsiflora	the	CR	Sonchus asper	the	R
Myosotis stricta	the	R	Sorbus aria	pha	C
Myosotis sylvatica	hem	C	Sorbus aucuparia	pha	C
Nanophyton erinaceum	cha	S	Sorbus dacica	pha	C
Neotinea ustulata	geo	C	Sorbus graeca	pha	CS
Nepeta cataria	hem	CR	Spiraea chamaedryfolia	pha	C
Nepeta nuda	hem	CR	Spiraea crenata	pha	CS
Nepeta ucranica	hem	CR	Spiraea hypericifolia	pha	CS
Noccaea kovatsii	hem	RS	Spiraea media	pha	C
Noccaea perfoliata	the	CRS	Stachys germanica	hem	C
Nonea pulla	hem	RS	Stachys officinalis	hem	C
Odontarrhena muralis	hem	CS	Stachys recta	hem	CS
Odontarrhena tortuosa aggr.	hem	CS	Sterigmotestum caspicum	cha	S
Odontites luteus	the	CRS	Stipa capillata	hem	C
Odontites vernus	the	CR	Stipa eriocaulis	hem	CS
Odontites vulgaris	the	CR	Stipa lessingiana	hem	CS
Onobrychis jaiiae	hem	CS	Stipa pennata	hem	CS
Onobrychis viciifolia aggr.	hem	CS	Stipa pulcherrima	hem	CS
Ononis pusilla	hem	CS	Stipa sareptana	hem	CS
Ononis spinosa	hem	CS	Stipa tirsia	hem	C
Onosma arenaria	hem	CS	Stipa zaleskii aggr.	hem	CS
Onosma heterophylla	hem	CS	Syringa vulgaris	pha	R
Onosma polyphylla	cha	S	Takhtajaniantha austriaca	hem	CS
Onosma rigida	hem	CS	Takhtajaniantha crispa	hem	CS
Onosma simplicissima	hem	CS	Takhtajaniantha pusilla	hem	S
Onosma staminea	cha	S	Tanacetum achilleifolium	hem	CS
Onosma tinctoria	hem	C	Tanacetum corymbosum	hem	C
Onosma visianii aggr.	hem	CS	Tanacetum kittaryanum	cha	CS
Ophrys insectifera	geo	C	Tanacetum millefolium	hem	CS
Orchis simia	geo	C	Tanacetum santolina	hem	CS
Origanum vulgare	hem	CR	Taraxacum sect. Dioszegia	hem	CR
Orlaya grandiflora	the	S	Taraxacum sect. Erythrocarpa	hem	CR
Ornithogalum collinum	geo	C	Taraxacum sect. Erythrosperma	hem	CR
Ornithogalum fischerianum	geo	C	Taraxacum sect. Macrocornuta	hem	CR
Ornithogalum pyrenaicum	geo	CS	Taraxacum sect. Piesis	hem	CR
Ornithogalum umbellatum	geo	C	Taraxacum sect. Scariosa	hem	CR
Orobanche alba	geo	CS	Taraxacum sect. Taraxacum	hem	CR
Orobanche caryophyllacea	geo	CS	Taraxacum sp.	hem	C
Orobanche elatior	geo	CS	Taxus baccata	the	C
Orobanche lutea	geo	CRS	Teesdalia nudicaulis	pha	RS
Orobanche sp.	geo	C	Tephrosia integrifolia	hem	CS
Orobanche teucrii	geo	CS	Teucrium botrys	the	RS
Otites sp.	hem	CRS	Teucrium chamaedrys	cha	CS
Oxalis acetosella	geo	C	Teucrium montanum	cha	CS
Oxytropis pilosa	hem	C	Teucrium polium	cha	CS
Paeonia tenuifolia	hem	CS	Thalictrum foetidum	hem	CS
Palmbia turgaica	hem	CS	Thalictrum minus	hem	C
Papaver dubium	the	RS	Thalictrum simplex	hem	C
Paronychia cephalotes	hem	CS	Thesium alpinum	hem	S
Pedicularis comosa	hem	S	Thesium brachyphyllum	hem	CS
Pedicularis interrupta	hem	S	Thesium linophyllum	hem	C
Pedicularis kaufmannii	hem	C	Thesium procumbens	hem	CS
Pedicularis physocalyx	cha	CS	Thesium ramosum	hem	CS
Pedicularis sibthorpii	hem	C	Thesium sp.	hem	C
Peltaria alliacea	hem	CRS	Thlaspi arvense	the	CR
Petrorhagia saxifraga	the	CRS	Thymelaea passerina	the	CR
Petrorhagia saxifraga	the	CRS	Thymus calcareus	cha	CS
Peucedanum austriacum	hem	CS	Thymus guberlinensis	cha	S
Peucedanum cervaria	hem	C	Thymus moldavicus	cha	S
Peucedanum oreoselinum	hem	C	Thymus pulegioides aggr.	cha	C
Peucedanum ruthenicum	hem	CS	Thymus serpyllum aggr.	cha	CS
Phegopteris connectilis	geo	CS	Thymus sp.	cha	C
Phleum montanum	hem	C	Thymus tauricus	pha	CS
Phleum phleoides	hem	C	Tilia cordata	cha	C
Phleum pratense	hem	C	Tilia platyphyllos	pha	C
Phlomis herba-venti subsp. pungens	hem	C	Tofieldia yaponica	hem	CS
Phlomis tuberosa	hem	C	Torilis italica	the	CR
Phyteuma orbiculare	hem	C	Torilis calyculata	hem	CRS
Phyteuma spicatum	hem	C	Tragopogon dubius	hem	CRS
Picea abies	pha	C	Tragopogon marginifolius	hem	S
Picris hieracioides	hem	RS	Tragopogon pratensis	hem	C
Pilosella cymosa	hem	CS	Tragopogon ruber	hem	S
Pilosella echioides	hem	CS	Trifolium alpestre	hem	C
Pilosella hoppeana	hem	C	Trifolium ambiguum	hem	C
Pilosella lactucella	hem	C	Trifolium arvense	the	CRS
Pilosella leptophyton	hem	C	Trifolium aureum	the	CR
Pilosella officinarum	hem	CS	Trifolium campestre	the	CR
Pilosella piloselloides aggr.	hem	CS	Trifolium dubium	the	CRS
Pilosella sp.	hem	C	Trifolium montanum	hem	C
Pimpinella saxifraga aggr.	hem	CS	Trifolium pratense	hem	C
Pinguicula alpina	hem	CS	Trifolium repens	hem	C
Pinus nigra	pha	C	Trifolium rubens	hem	C
Pinus sylvestris	pha	C	Trigonella gladiata	the	R
Piptatherum holciforme	hem	CS	Trinia glauca	hem	CS
Piptatherum virescens	hem	C	Trinia multicaulis	hem	C
Plantago argentea	hem	CS	Trisetum flavescens aggr.	hem	C
Plantago lanceolata	hem	C	Tulipa biflora	geo	CS
Plantago maritima subsp. ciliata	hem	CS	Tulipa sp.	geo	CS
Plantago media	hem	C	Tulipa suaveolens	geo	CS
Plantago tenuiflora	the	CRS	Tulipa sylvestris	geo	C
Platanthera bifolia	geo	C	Turritis glabra	hem	CR
Poa alpina	hem	CS	Tussilago farfara	geo	CRS
Poa angustifolia	hem	C	Tulipa scythica	geo	C
Poa badensis	hem	CS	Ulmus minor	pha	C
Poa bulbosa	hem	RS	Ulmus sp.	pha	C
Poa compressa	hem	RS	Urtica dioica	hem	CR
Poa nemorinii	hem	CS	Vaccinium montanum	cha	CS
Poa nivalis	hem	C	Valeriana officinalis	hem	C
Poa pannonica	hem	C	Valeriana officinalis aggr.	hem	C
Poa pratensis	hem	C	Valeriana stolonifera	hem	C
Poa sterilis	hem	CS	Valeriana tripteris	hem	CS
Poa versicolor	hem	CS	Valerianella locusta aggr.	the	R
Polycnemum arvense	the	CRS	Valenata dubia	the	RS
Polygala amara	hem	C	Verbascum chaixii	hem	CR
Polygala comosa	hem				

Supplement E3. Algorithm of the Grime's triangle construction.

Anhang E3 Algorithmus zur Konstruktion des Grime-Dreiecks.

1. Calculating the indices of CRS using the formulas:

$$Ss = S + (CS/2) + (SR/2) + (CRS/3); Cc = C + (CS/2) + (CR/2) + (CRS/3); Rr = R + (CR/2) + (RS/2) + (CRS/3)$$

S is the sum of species with expressed stress-tolerant strategies, C is the sum of species with expressed concurrent strategies, and R is the sum of species with expressed ruderal strategies. The sum values are considered in full.

CS is the sum of simultaneous competitors and stress-tolerant species, CR is the sum of simultaneous competitors and ruderal species, SR is the sum of simultaneous stress-tolerant and ruderal species, and RS is the sum of simultaneous ruderal and stress-tolerant species. The sum values are divided into two.

CRS is the sum of species more or less balanced features of all three strategy types. The sum values are divided into three.

Ss is the sum of all species exhibiting traits of the stress-tolerant strategy, Cc is the sum of all species exhibiting traits of the competitive strategy, and Rr is the sum of all species exhibiting traits of the ruderal strategy.

2. Calculating the ratios between the indicators: Ss: Cc, Rr: Cc, Ss: Rr, proportions of C: S: R = 1.0: X: Y, where X and Y are the proportions of S and R relative to C=1.

3. Displaying the indicators (in percentages) of species ratios on the sides of the Grime triangle, where the vertices correspond to 100%:

- On the C–S side, the point for Cc = $100 \cdot Cc / (Cc + Ss)$, and thus the position for Ss = $100 - Cc$.
- On the C–R side, the position for Rr = $100 \cdot Rr / (Rr + Cc)$, and thus the point for Cc = $100 - Rr$.
- On the S–R side, the position for Ss = $100 \cdot Ss / (Ss + Rr)$, and thus the point for Rr = $100 - Ss$.

The points are plotted on the sides of the triangle and connected. Such a representation becomes an object of further analysis, which we illustrated with specific results.

4. Determining the "centre of mass" of the triangle based on the mean values of the indicators. The "centre" corresponds to point N (x, y), whose coordinates are located at the intersection of its medians. As is known, the point where the medians intersect divides each median in a ratio of 2:1, starting from the vertex. For this, the indicators of the vertices are projected onto the X-axis (coinciding with the lower side of the triangle) and the Y-axis, which are assigned corresponding values. Then, the position of the "centre of mass" is calculated using the formulas:

- $x = (x_1 + x_2 + x_3) / 3$
- $y = (y_1 + y_2 + y_3) / 3$

These points can be constructed graphically. The obtained positions are overlaid onto the Grime triangle, which shows nine fields and a complex transitional zone configuration (CRS). Additionally, fields are highlighted where descriptions cannot fall (horizontal hatching).

Supplement E4. Full synoptic table showing the percentage constancy and fidelity of species of the six clusters. Diagnostic species are ordered by decreasing fidelity measured with the phi-coefficient. Species with a phi-value greater than 0.3 are considered diagnostic and shaded by light grey colour; species with a phi-value greater than 0.5 are considered highly diagnostic and shaded by dark grey colour. Species that are highly diagnostic or diagnostic in one cluster are accepted as diagnostic for associations; species that are highly diagnostic or diagnostic in more than one cluster are accepted as diagnostic for high-rank syntaxa

Anhang E4. Vollständige synoptische Tabelle mit der prozentualen Steitigkeit und Treue der Arten der sechs Cluster. Diagnostische Arten sind nach abnehmender Treue, gemessen am *Phi*-Koeffizienten, sortiert. Arten mit einem Phi-Wert über 0.3 gelten als diagnostisch und sind hellgrau hinterlegt; Arten mit einem *Phi*-Wert über 0,5 gelten als hochdiagnostisch und sind dunkelgrau hinterlegt. Arten, die in einem Cluster hochdiagnostisch oder diagnostisch sind, gelten als diagnostisch für Assoziationen; Arten, die in mehr als einem Cluster hochdiagnostisch oder diagnostisch sind, gelten als diagnostisch für hochrangige Syntaxa.

Number of clusters	1	2	3	4	5	6	7	8	9	1	11	12	13
Number of relevés	35	11	73	243	92	280	116	290	196	61	111	80	55
AB. <i>Anabasis cretaceae</i> - <i>Agropyron desertori</i>													
<i>Astragalus kustanaiicus</i>	63	.	.	.	.	.	.	.	.	.	.	.	.
<i>Bassia prostrata</i>	89	9	19	5	2	2	.	.	.	.	.	1	.
<i>Stierigmostemum caspicum</i>	69	9	7	.	.	.	.	.	.	.	.	.	.
<i>Galatella tatarica</i>	57	.	4	.	.	.	.	.	.	.	.	.	.
<i>Krascheninnikovia ceratoides</i>	63	.	7	5	3	1	.	.	.	.	.	.	.
<i>Tulipa scythica</i>	49	.	5	.	.	.	.	.	.	.	.	.	.
<i>Lappula microcarpa</i>	63	.	27	.	.	.	.	.	.	.	.	.	.
<i>Allium tulipifolium</i>	43	.	7	.	.	.	.	.	.	.	.	.	.
<i>Tulipa biflora</i>	37	.	1	.	.	.	.	.	.	.	.	.	.
<i>Tulipa sylvestris</i>	37	.	1	.	.	.	.	.	.	.	.	.	.
<i>Tragopogon ruber</i>	40	.	5	.	.	.	.	.	.	.	.	.	.
<i>Taraxacum</i> sect. <i>Macrocornuta</i>	31	.	1	.	.	.	.	.	.	.	.	.	.
<i>Isatis gymnocarpa</i>	29	.	.	.	.	.	.	.	.	.	.	.	.
<i>Meniocus linifolius</i>	37	.	10	.	1	.	.	.	.	.	.	.	.
<i>Asparagus ierdiensis</i>	34	.	7	.	.	.	.	.	.	.	.	.	.
<i>Stipa sareptana</i>	46	18	12	.	.	.	.	.	.	.	.	.	.
<i>Litwinovia tenuissima</i>	23	.	.	.	.	.	.	.	.	.	.	.	.
<i>Caragana balchashensis</i>	23	.	1	.	.	.	.	.	.	.	.	.	.
<i>Poa bulbosa</i>	40	.	4	1	.	5	7	.	1	.	5	6	.
<i>Palmbia turgaica</i>	23	.	3	.	.	.	.	.	.	.	.	.	.
<i>Ferula caspica</i>	23	.	3	.	.	.	.	.	.	.	.	.	.
<i>Rindera tetraspis</i>	31	18	.	.	.	.	.	.	.	.	.	.	.
<i>Artemisia semiarida</i>	31	18	3	.	.	.	.	.	.	.	.	.	.
<i>Gagea bulbifera</i>	17	.	.	.	.	.	.	.	.	.	.	.	.
<i>Allium lineare</i>	17	.	.	.	.	.	.	.	.	.	.	.	.
<i>Pseudoscedum lievenii</i>	17	.	.	.	.	.	.	.	.	.	.	.	.
<i>Limonium meyeri</i>	17	.	.	.	.	.	.	.	.	.	.	.	.
<i>Rochelia disperma</i> subsp. <i>retorta</i>	17	.	.	.	.	.	.	.	.	.	.	.	.
<i>Anabasis salsa</i>	23	9	.	.	.	.	.	.	.	.	.	.	.
<i>Artemisia lessingiana</i>	29	.	21	.	.	.	.	.	.	.	.	.	.
<i>Astragalus temirensis</i>	14	.	.	.	.	.	.	.	.	.	.	.	.
<i>Tulipa suaveolens</i>	14	.	.	.	.	.	.	.	.	.	.	.	.
<i>Filago arvensis</i>	14	.	.	.	.	.	.	.	.	.	.	.	.
<i>Eriosynaphe longifolia</i>	14	.	.	.	.	.	.	.	.	.	.	.	.
<i>Klasea erucifolia</i>	14	.	.	.	.	.	.	.	.	.	.	.	.
<i>Linaria incompleta</i>	20	.	7	.	.	.	.	.	.	.	.	.	.
<i>Artemisia lerchiana</i>	37	18	33	2	.	1	.	.	.	.	.	.	.
<i>Androsace maxima</i>	17	.	5	.	.	.	.	.	.	.	.	.	.
<i>Takhtajaniantha pusilla</i>	11	.	.	.	.	.	.	.	.	.	.	.	.
<i>Allium ierdiense</i>	11	.	.	.	.	.	.	.	.	.	.	.	.
<i>Silene viscosa</i>	11	.	.	.	.	.	.	.	.	.	.	.	.
<i>Isotrlion tataricum</i>	11	.	.	.	.	.	.	.	.	.	.	.	.
<i>Ornithogalum fischerianum</i>	11	.	.	.	.	1	.	.	.	.	.	.	.
<i>Allium saxatile</i>	14	.	4	.	.	.	.	.	.	.	.	.	.
<i>Artemisia nitrosa</i>	14	.	4	.	.	.	.	.	.	.	.	.	.
<i>Astragalus rupifragus</i>	11	.	1	.	.	.	.	.	.	.	.	.	.
AB. <i>Sileno fruticosae</i> - <i>Nanophyton erinacei</i>													
<i>Matthiola tatarica</i>	100	.	.	.	.	.	.	.	.	.	.	.	.
<i>Gypsophila diffusa</i>	100	.	.	.	.	.	.	.	.	.	.	.	.
<i>Onosma staminea</i>	91	.	.	.	.	.	.	.	.	.	.	.	.
<i>Nanophyton erinaceum</i>	11	91	18	.	.	.	.	.	.	.	.	.	.
<i>Atrophaxis spinosa</i>	.	64	.	.	.	.	.	.	.	.	.	.	.
<i>Atrophaxis replicata</i>	3	64	.	.	.	.	.	.	.	.	.	.	.
<i>Artemisia terrae-albae</i>	3	64	.	.	.	.	.	.	.	.	.	.	.
<i>Galium humifusum</i>	.	64	.	1	.	3	.	1	1	.	3	.	.
<i>Lepidium meyeri</i>	.	73	25	4	.	.	.	.	.	.	.	.	.
<i>Silene antri-jovis</i>	.	55	4	.	.	.	.	.	.	.	.	.	.
<i>Lomelosia isetensis</i>	6	73	25	.	.	.	.	.	.	.	.	.	.
<i>Astragalus medius</i>	3	36	1	.	.	.	.	.	.	.	.	.	.
<i>Ephedra distachya</i>	34	73	47	.	9	3	.	.	.	.	.	.	.
<i>Astragalus stenoceras</i>	.	27	.	.	.	.	.	.	.	.	.	.	.
<i>Rhammatophyllum pachyrhizum</i>	.	27	1	.	.	.	.	.	.	.	.	.	.
<i>Limonium suffruticosum</i>	3	27	.	.	.	.	.	.	.	.	.	.	.
<i>Convolvulus fruticosus</i>	.	18	.	.	.	.	.	.	.	.	.	.	.
<i>Echinops ritro</i>	23	45	41	5	4	14	.	.	9	.	1	.	.
AB. <i>Anthemido trozkianae</i> - <i>Artemision salsoloides</i>													
<i>Archanthemis trozkiana</i>	6	.	90	.	.	.	.	.	.	.	.	.	.
<i>Zygophyllum pinnatum</i>	.	9	67	.	.	.	.	.	.	.	.	.	.
<i>Atrophaxis decipiens</i>	.	.	48	.	.	.	.	.	.	.	.	.	.
<i>Astragalus tenuifolius</i>	23	.	56	.	.	.	.	.	.	.	.	.	.
<i>Hedysarum tscherkassovae</i>	3	.	36	.	.	.	.	.	.	.	.	.	.
<i>Jurinea kirghisorum</i>	.	.	25	.	.	.	.	.	.	.	.	.	.
<i>Gypsophila rupestris</i>	.	.	21	.	.	.	.	.	.	.	.	.	.
<i>Hedysarum razoumouianum</i>	3	.	18	.	.	.	.	.	.	.	.	.	.
<i>Galium octonarum</i>	9	.	27	9	1	5	.	.	.	.	.	.	.
<i>Gypsophila altissima</i>	.	.	12	.	.	.	.	.	.	.	.	.	.
<i>Atrophaxis frutescens</i>	.	.	14	2	.	.	.	.	.	.	.	.	.
<i>Crambe tatarica</i> aggr.	11	.	22	3	.	2	.	.	.	.	.	.	.
<i>Psephellus marschallianus</i> aggr.	34	.	36	5	34	34	.	2	.	1	1	.	.
AB. <i>Sileno supinae</i> - <i>Artemision hololeuca</i>													
<i>Hyssopus officinalis</i>	.	.	.	81	8	10	.	.	.	.	.	.	.
<i>Scrophularia cretacea</i>	.	.	.	52	13	6	.	.	.	.	.	.	.
<i>Artemisia hololeuca</i>	.	.	.	48	4	12	.	.	.	.	.	.	.
<i>Festuca cretacea</i>	.	.	.	18	1	1	.	.	.	.	.	.	.
<i>Plantago maritima</i> subsp. <i>ciliata</i>	.	.	.	16	.	6	.	.	.	.	.	.	.
AB. <i>Euphorbia cretophilae</i> - <i>Thymion cretaei</i>													
<i>Euphorbia petrophila</i>	.	.	.	7	84	5	10	.	.	.	.	1	.
<i>Jurinea stoehadifolia</i>	.	.	.	.	73	7	1	.	3	.	.	.	.
<i>Hedysarum grandiflorum</i>	.	.	1	2	32	10	.	.	.	.	.	.	.
<i>Reseda lutea</i>	.	.	1	23	55	38	14	1	1	.	4	1	.
<i>Genista scythica</i>	.	.	.	.	20	1	.	.	.	.	.	.	.
<i>Onosma simplicissima</i>	3	.	11	14	49	38	.	.	.	.	.	.	.
<i>Odontarrhena tortuosa</i> aggr.	6	.	33	2	38	9	14	.	.	.	.	.	.
<i>Rhaponticoides talievii</i>	.	.	.	.	11	.	.	.	.	.	.	.	.
AB. <i>Centaureo carbonatae</i> - <i>Koelerion talievii</i>													
<i>Bromopsis riparia</i>	.	.	.	2	8	35	.	3	1	.	.	.	.
<i>Polygala sibirica</i>	.	.	.	6	1	33	.	2	.	1	1	.	.
<i>Viola ambigua</i>	.	.	.	1	5	33	3	.	1	.	3	.	.
<i>Koeleria talievii</i>	.	.	.	7	1	34	.	.	.	.	.	.	.
<i>Stipa pennata</i>	.	.	.	2	1	33	.	1	9	.	2	3	.
<i>Salvia nutans</i>	.	.	.	1	29	39	.	4	.	1	1	.	.
<i>Euphorbia seguieriana</i>	34	9	10	25	8	64	27	.	4	.	9	.	.
<i>Polygala cretacea</i>	.	.	1	26	9	36	.	.	.	.	.	.	.
<i>Astragalus albicaulis</i>	.	.	.	23	13	32	.	.	1	.	.	.	.
<i>Astragalus austriacus</i>	.	.	1	1	1	20	.	.	5	.	1	.	.
<i>Euphorbia nicaensis</i>	.	.	.	9	3	21	1	.	3	.	3	.	.
Seseli leucospermum-Fumana procumbens grasslands													
<i>Seseli leucospermum</i>	.	.	.	.	.	44	.	.	.	1	.	.	.
<i>Fumana procumbens</i>	.	.	.	.	1	.	65	.	19	.	11	.	5
<i>Paronychia cephalotes</i>	.	.	.	.	3	.	41	2	.	5	.	.	.
<i>Bromopsis cappadocica</i>	.	.	1	2	1	34	.	.	.	.	.	.	.
<i>Allium moschatum</i>	.	.	.	.	.	.	28	.	.	1	.	.	.
<i>Mimuartia setacea</i>	.	.	.	7	.	.	58	5	1	.	17	20	5
<i>Alyssum montanum</i> aggr.	.	.	.	.	4	50	5	16	.	7	8	.	.
<i>Thymus tauricus</i>	.	.	.	.	.	.	24	.	.	.	.	.	.
<i>Genista albidia</i>	.	.	.	2	.	.	26	.	.	.	.	.	.
<i>Asperula supina</i>	.	.	1	1	1	26	.	.	.	.	.	.	.
<i>Helianthemum canum</i> aggr.	.	.	4	38	14	73	13	22	.	11	25	.	.
<i>Takhtajaniantha crispa</i>	.	.	.	1	.	.	22	.	.	.	.	.	.
<i>Carex liparocarpus</i>	.	.	.	.	.	.	20	.	.	.	.	.	.
<i>Odontites luteus</i>	.	.	13	16	6	47	1	.	14	8	.	.	.
<i>Sideritis taurica</i>	.	.	.	1	.	.	19	.	.	.	.	.	.
<i>Chrysopogon gryllus</i>	.	.	.	.	.	.	24	2	.	6	.	.	.
<i>Stipa eriocalis</i>	.	.	.	.	.	.	19	2	1	.	1	.	.
<i>Anthyllis vulneraria</i> aggr.	.	.	.	1	.	.	56	7	38	.	16	13	42
<i>Stipa pulcherrima</i>	.	.	.	1	8	31	31	2	8	2	5	6	.
<i>Cerastium semidecandrum</i>	.	.	.	.	.	.	20	.	.	6	.	2	.
<i>Veronica taurica</i>	.	.	.	.	.	.	13	.	.	.	.	.	.
<i>Iberis saxatilis</i>	.	.	.	.	.	.	13	.	.	.	.	.	.
<i>Draba lasiocarpa</i>	.	.	.	.	.	.	26	1	10	.	5	.	5
<i>Takhtajaniantha austriaca</i>	.	.	.	.	.	.	30	1	26	.	3	1	4
<i>Convolvulus calvertii</i> subsp. <i>calvertii</i>	.	.	.	.	.	.	12	.	.	.	.	.	.
<i>Koeleria brevis</i> aggr.	.	.	.	1	1	14	.	.	.	.	.	.	.
<i>Cruciata taurica</i>	.	.	.	1	.	.	13	.	.	.	.	.	.
<i>Potentilla astracanica</i>	.	.	.	.	1	.	13	.	.	.	.	.	.
<i>Linum tenuifolium</i>	.	.	5	25	4	44	2	38	.	7	4	2	.
<i>Potentilla recta</i>	.	.	.	2	3	22	8	.	.	.	1	2	.

ID clusters	1	2	3	4	5	6	7	8	9	1	11	12	13
Number of relevés	35	11	73	243	92	280	116	290	196	61	111	80	55
Other taxa occurred in the communities (continued)													
<i>Corylus avellana</i>	.	.	.	.	.	.	.	1	1	.	.	.	4
<i>Cota tinctoria</i>	.	.	2	7	2	1	4	.	3	1	1	2	.
<i>Cotinus coggygria</i>	.	.	.	2	.	.	.	3	2	3	1	1	.
<i>Cotoneaster integrerrimus</i> aggr.	.	.	.	.	.	5	8	8	13	.	5	9	.
<i>Cotoneaster</i> sp.	.	.	.	.	.	.	.	.	.	.	.	2	.
<i>Cousinia astracanica</i>	.	1	.	.	.	.	.	.	.	.	.	.	.
<i>Crataegus laevigata</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Crataegus monogyna</i>	.	.	.	.	.	3	1	1	.	4	4	2	.
<i>Crataegus rhipidophylla</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Crepis foetida</i>	.	.	1	.	1	.	1	.	.	1	.	.	.
<i>Crepis jacquini</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Crepis neglecta</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Cruciatia glabra</i>	.	.	.	.	.	.	3	1	.	.	2	.	.
<i>Crupina vulgaris</i>	.	.	.	.	1	.	.	.	4	.	.	.	.
<i>Ctenidium molluscum</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Cuscuta campestris</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Cuscuta epithymum</i> aggr.	.	1	.	.	2	.	1	3	3	4	.	.	.
<i>Cuscuta europaea</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Cuscuta</i> sp.	.	1	.	1	.	.	.	1	.	.	.	.	.
<i>Cyanus triumfetti</i>	.	.	.	.	.	.	13	15	.	8	.	4	.
<i>Cynodon dactylon</i>	.	.	.	.	.	3	.	.	.	.	.	.	.
<i>Cynoglossis barrelieri</i>	.	.	.	.	.	.	3	.	.	.	1	.	.
<i>Cynosurus cristatus</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Cystopteris fragilis</i>	.	.	.	.	.	.	6	15	.	1	2	.	.
<i>Cytisus albus</i>	.	.	.	.	.	.	6	2	.	2	8	.	.
<i>Cytisus austriacus</i>	.	.	.	1	.	1	2	.	2	.	.	.	.
<i>Cytisus blockianus</i>	.	.	.	.	.	.	.	.	.	.	3	.	.
<i>Cytisus granticus</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Cytisus hirsutus</i> aggr.	.	.	1	5	8	13	10	6	3	5	4	2	.
<i>Cytisus paezcoskii</i>	.	.	.	.	.	.	.	2	.	.	3	.	.
<i>Cytisus podolicus</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Cytisus procumbens</i>	.	.	.	.	.	.	2	.	1	.	.	.	.
<i>Cytisus</i> sp.	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Dactylis glomerata</i>	.	.	.	.	.	.	1	.	.	.	2	.	.
<i>Dactylorhiza sambucina</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Danthonia alpina</i>	.	.	.	.	.	.	2	1	.	.	.	.	.
<i>Daphne cneorum</i>	.	.	.	.	.	.	6	.	.	.	.	.	.
<i>Daucus carota</i>	.	2	.	1	.	1	.	.	3	4	.	.	.
<i>Deschampsia cespitosa</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Descraineria sophia</i>	3	.	1	.	.	.	.	.	.	.	.	.	.
<i>Dianthus carthusianorum</i> aggr.	.	.	.	.	.	12	22	11	5	21	4	13	.
<i>Dianthus giganteus</i>	.	.	.	.	.	2	1	.	.	6	.	.	.
<i>Dianthus henteri</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Dianthus lanceolatus</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Dianthus nitidus</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Dianthus plumarius</i>	.	.	.	.	.	7	1	3	.	.	.	.	.
<i>Dianthus praecox</i>	.	.	.	.	.	.	.	14	.	5	.	9	.
<i>Dianthus pseudarmeria</i>	.	.	.	4	1	.	.	.	.	.	.	.	.
<i>Dianthus ramosissimus</i>	3	.	.	.	.	.	.	.	.	.	.	.	.
<i>Dichoropetalum carvifolia</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Dicranella heteromalla</i>	.	.	.	.	.	.	.	.	.	.	2	.	.
<i>Dicranum boreale</i>	.	.	.	.	.	.	.	.	.	3	.	.	.
<i>Dicranum scoparium</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Dictamnus albus</i> aggr.	.	.	1	.	.	4	1	5	6	1	.	.	.
<i>Didymodon acutus</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Didymodon</i> sp.	.	.	.	.	.	.	.	.	.	.	1	.	.
<i>Digitalis grandiflora</i>	.	.	.	.	.	.	13	2	.	3	.	2	.
<i>Diploschistes muscorum</i>	.	.	.	.	.	.	.	.	.	.	5	.	.
<i>Diplotaxis cretacea</i>	.	.	5	11	4	.	.	.	.	.	.	.	.
<i>Diplotaxis muralis</i>	.	.	.	.	1	.	.	1	.	.	1	.	.
<i>Diplotaxis tenuifolia</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Doronicum columnae</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Dorycnium pentaphyllum</i>	.	.	.	.	.	10	1	19	.	8	.	.	.
<i>Draba aizoides</i>	.	.	.	.	.	.	.	1	.	.	4	.	.
<i>Draba cuspidata</i>	.	.	.	.	.	6	.	.	.	.	.	.	.
<i>Draba nemorosa</i>	.	.	.	.	.	.	.	.	2	.	.	.	.
<i>Draba podolica</i>	.	.	.	1	.	.	.	1	.	3	.	.	.
<i>Draba verna</i>	.	.	.	.	1	.	.	2	5	8	.	2	.
<i>Dracocephalum austriacum</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Drymocalis geoides</i>	.	.	.	.	.	3	.	.	.	.	.	.	.
<i>Echinops bannaticus</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Echinops sphacelcephalus</i>	.	.	.	.	.	.	1	.	.	.	3	.	.
<i>Echium vulgare</i>	.	.	.	.	8	2	7	8	3	23	18	18	.
<i>Eclatanthus graminifolius</i>	.	.	.	.	.	.	.	.	.	.	.	.	.
<i>Elymus caninus</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Elytrigia caespitosa</i> subsp. <i>nodosa</i>	.	.	.	.	.	2	.	.	.	.	.	.	.
<i>Elytrigia intermedia</i>	.	.	.	1	10	.	4	4	.	11	18	.	.
<i>Elytrigia repens</i>	1	1	.	9	3	1	.	2	1	.	.	.	.
<i>Elytrigia stipifolia</i>	.	.	2	2	3	.	.	.	.	.	.	.	.
<i>Elytrigia strigosa</i>	.	.	.	.	.	9	.	.	.	.	.	.	.
<i>Encalypta vulgaris</i>	.	.	.	.	.	.	1	3	8	9	.	.	.
<i>Enchylum bachmantianum</i>	.	.	.	.	.	.	.	.	1	.	.	.	.
<i>Enchylum tenax</i>	.	2	.	3	.	.	.	.	.	9	.	.	.
<i>Epipactis atrorubens</i>	.	.	.	.	.	.	1	9	.	1	.	.	.
<i>Eremopyrum triticeum</i>	3	.	.	.	.	.	.	.	.	.	.	.	.
<i>Eremurus spectabilis</i>	.	.	.	1	.	.	.	.	.	.	.	.	.
<i>Erigeron acris</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Erigeron canadensis</i>	.	.	.	1	.	.	.	.	.	.	.	.	.
<i>Erodium cicutarium</i>	.	.	.	.	.	.	.	.	5	1	.	.	.
<i>Erucastrum nasturtifolium</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Eryngium planum</i>	.	.	.	2	1	.	.	.	.	1	.	.	.
<i>Erysimum canum</i>	.	1	1	.	.	.	.	.	.	.	.	.	.
<i>Erysimum comatum</i>	.	.	.	.	.	.	1	.	1	.	.	.	.
<i>Erysimum cretaceum</i>	.	.	.	1	.	.	.	.	.	.	.	.	.
<i>Erysimum cuspidatum</i>	.	.	.	.	.	6	.	.	.	.	.	.	.
<i>Erysimum leptostylum</i>	.	1	3	1	.	.	.	.	.	.	.	.	.
<i>Erysimum odoratum</i>	.	.	.	.	.	9	30	14	3	30	13	24	.
<i>Erysimum rhaeticum</i>	.	.	.	.	.	.	.	.	.	.	.	.	.
<i>Erysimum</i> sp.	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Erysimum taljevii</i>	.	1	.	1	.	.	.	.	.	.	.	.	.
<i>Erysimum ucrainicum</i>	.	8	.	.	.	.	.	.	.	.	.	.	.
<i>Erysimum wilmannii</i>	.	.	.	.	.	8	10	.	.	13	.	.	.
<i>Euonymus verrucosus</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Euphorbia agraria</i>	.	.	.	1	10	.	2	1	.	.	.	.	.
<i>Euphorbia amygdaloides</i>	.	.	.	.	.	.	1	1	.	2	.	2	.
<i>Euphorbia angulata</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Euphorbia epithymoides</i>	.	.	.	.	.	2	5	2	4	.	.	.	.
<i>Euphorbia esula</i>	3	.	.	1	.	.	2	.	.	.	.	.	.
<i>Euphorbia myrsinites</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Euphorbia semivillosa</i>	.	1	.	1	.	.	.	.	.	.	.	.	.
<i>Euphorbia stricta</i>	.	.	.	.	.	.	.	.	.	9	4	.	.
<i>Euphrasia hirtella</i>	.	1	.	3	.	.	.	1	4	.	2	.	.
<i>Euphrasia rostkoviana</i>	.	.	.	.	.	.	3	4	.	1	4	.	.
<i>Euphrasia salisburgensis</i>	.	.	.	.	.	.	.	.	.	.	1	2	.
<i>Euphrasia stricta</i> aggr.	.	1	.	8	.	1	4	3	1	.	5	.	.
<i>Euphrasia taurica</i>	.	.	.	.	2	.	.	.	.	.	.	.	.
<i>Evernia mesomorpha</i>	.	.	.	.	.	.	1	1	.	3	.	.	.
<i>Fagus sylvatica</i>	.	.	.	.	.	.	1	1	.	.	.	.	.
<i>Falcaria vulgaris</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Fallopia convolvulus</i>	.	.	.	.	.	1	.	1	2	3	.	4	.
<i>Fallopia dumetorum</i>	.	.	.	.	.	1	.	2	1	.	.	.	.
<i>Ferula tatarica</i>	11	.	3	.	.	.	.	.	.	.	.	.	.
<i>Ferulago campestris</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Ferulago sylvatica</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Ferulago taurica</i>	.	.	.	.	.	5	.	.	.	.	.	.	.
<i>Festuca glauca</i>	.	.	.	.	.	.	6	.	.	.	.	.	.
<i>Festuca heterophylla</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Festuca rubra</i>	.	.	.	1	.	1	1	.	.	4	.	.	.
<i>Filago germanica</i>	.	.	.	.	2	.	.	.	.	.	.	.	.
<i>Filipendula vulgaris</i>	.	.	.	4	17	1	.	13	5	1	5	.	.
<i>Fissidens taxifolius</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Flexitrichum flexicaule</i>	.	.	.	.	.	1	2	.	1	8	7	.	.
<i>Fragaria moschata</i>	.	.	.	.	.	7	.	10	.	.	15	.	.
<i>Fragula alnus</i>	.	.	.	.	.	1	.	.	1	.	.	.	.
<i>Fraxinus excelsior</i>	.	.	.	.	.	3	.	.	.	.	.	.	.
<i>Fraxinus lanceolata</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Fraxinus ornus</i>	.	.	.	.	.	8	.	.	4	.	.	.	.
<i>Fritillaria orientalis</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Fumaria officinalis</i>	.	.	.	.	.	.	.	2	.	.	.	.	.
<i>Gagea pratensis</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Gagea pusilla</i>	3	.	.	.	.	.	.	.	1	.	.	.	.
<i>Galanthus nivalis</i>	.	.	.	.	.	.	.	.	1	.	.	.	.
<i>Galatella angustissima</i>	.	1	.	.	.	.	.	.	.	.	.	.	.
<i>Galatella linosyris</i>	.	.	.	1	7	.	1	.	3	.	.	.	.
<i>Galatella villosa</i>	14	.	3	.	1	4	.	.	.	.	.	.	.
<i>Galeopsis angustifolia</i>	.	.	.	.	.	.	1	.	.	.	2	.	.
<i>Galeopsis ladanum</i>	.	.	.	.	.	.	.	2	.	.	.	.	.
<i>Galitzkya sphathulata</i>	.	8	.	.	.	.	.	.	.	.	.	.	.
<i>Galium anisophyllum</i>	.	.	.	.	.	.	1	1	.	.	.	.	.
<i>Galium aparine</i>	.	.	.	.	.	.	.	2	.	.	.	.	.
<i>Galium baillonii</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Galium biebersteinii</i>	.	.	.	.	3	5	.	.	.	.	.	.	.
<i>Galium boreale</i>	.	.	.	.	1	.	.	1	.	5	.	.	.
<i>Galium glaucum</i>	.	.	.	.	1	13	1	10	5	5	23	9	.
<i>Galium pumilum</i> aggr.	.	.	.	.	.	.	12	.	2	.	.	.	.
<i>Galium transcarpaticum</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Galium verum</i> aggr.	.	5	3	12	13	7	3	2	4	9	5	.	.
<i>Galium volynicum</i>	.	.	.	.	.	.	1	.	1	.	.	.	.
<i>Gehebia juliae</i>													

ID clusters	1	2	3	4	5	6	7	8	9	1	11	12	13
Number of relevés	35	11	73	243	92	280	116	290	196	61	111	80	55
Other taxa occurred in the communities (continued)													
<i>Salsola tragus</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Salvia nemorosa</i>	.	.	.	.	.	3	.	.	.	.	.	4	.
<i>Salvia pratensis</i> aggr.	.	.	.	.	.	1	3	2	14	.	10	8	4
<i>Salvia scabiosifolia</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Salvia verbenaca</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Salvia verticillata</i>	.	.	.	3	11	18	.	13	11	.	2	8	25
<i>Sanguisorba minor</i>	.	.	.	.	.	4	51	12	49	.	50	3	35
<i>Sanguisorba officinalis</i>	.	.	.	.	.	.	.	.	.	.	.	.	4
<i>Saponaria bellidifolia</i>	.	.	.	.	.	.	.	6	1	.	.	4	.
<i>Saponaria glutinosa</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Satureja montana</i> aggr.	.	.	.	.	2	.	4	1	.	.	.	.	.
<i>Saxifraga adscendens</i>	.	.	.	.	.	.	.	2	.	.	.	.	.
<i>Saxifraga marginata</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Saxifraga paniculata</i>	.	.	.	.	.	.	.	22	2	20	1	.	16
<i>Saxifraga tridactylites</i>	.	.	.	.	.	.	.	2	3	7	5	.	9
<i>Scabiosa canescens</i>	.	.	.	.	.	.	16	.	7	.	2	.	.
<i>Scabiosa columbaria</i>	.	.	.	.	1	.	3	8	3	.	1	5	.
<i>Scabiosa lucida</i>	.	.	.	.	.	.	.	4	2	.	.	.	.
<i>Schedonorus arundinaceus</i>	.	.	.	1	.	3	.	.	.	.	.	.	.
<i>Schedonorus pratensis</i>	.	.	.	.	.	1	.	.	.	.	1	.	2
<i>Schistidium apocarpum</i>	.	.	.	.	.	1	.	.	.	.	.	5	.
<i>Schistidium brunnescens</i>	.	.	.	.	.	.	.	.	.	.	.	1	.
<i>Schistidium</i> sp.	.	.	.	.	.	.	.	.	.	.	.	1	.
<i>Scleranthus annuus</i>	.	.	.	.	.	.	.	1	.	3	.	.	.
<i>Scleranthus perennis</i>	.	.	.	.	.	.	.	.	.	.	.	.	2
<i>Scorzonera laciniata</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Scorzonera mollis</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Scorzonera purpurea</i>	.	.	.	.	.	.	.	1	2	.	1	3	.
<i>Scorzoneroides autumnalis</i>	.	.	.	.	.	.	.	1	3	.	.	.	5
<i>Scrophularia heterophylla</i> subsp. <i>laciniata</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Scrophularia rupestris</i>	.	.	.	.	1	.	1	3	.	.	.	.	.
<i>Scutellaria albidia</i>	.	.	.	1	.	1	.	.	.	.	.	.	.
<i>Scutellaria orientalis</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Scutellaria supina</i>	.	.	.	.	5	4	.	.	.	.	.	.	.
<i>Scytinium lichenoides</i>	.	.	.	.	.	.	.	.	.	.	.	.	4
<i>Securigera elegans</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Securigera varia</i>	.	.	.	4	9	31	1	18	5	.	8	11	16
<i>Sedum acre</i>	.	.	.	.	.	1	11	5	38	32	44	44	.
<i>Sedum annuum</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Sedum rubens</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Sedum rupestre</i>	.	.	.	.	.	.	.	.	.	3	.	.	.
<i>Selaginella helvetica</i>	.	.	.	.	.	.	.	6	.	.	.	.	.
<i>Senpervivum marmoreum</i>	.	.	.	.	.	.	.	20	1	16	7	1	.
<i>Senpervivum ruthenicum</i>	.	.	.	.	.	.	.	.	.	.	.	6	.
<i>Senpervivum tectorum</i>	.	.	.	.	.	.	.	1	.	.	1	.	.
<i>Senecio glaucus</i> subsp. <i>coronopifolius</i>	.	.	1	.	.	.	.	.	.	.	.	.	.
<i>Senecio leucanthemifolius</i> subsp. <i>vernalis</i>	.	.	.	.	1	.	.	.	.	.	.	1	2
<i>Senecio</i> sp.	.	.	.	.	.	.	9	.	.	.	.	.	.
<i>Senecio squalidus</i> aggr.	.	.	.	.	.	.	.	3	.	.	.	1	.
<i>Senecio umbrosus</i>	.	.	.	.	.	.	.	.	3	.	.	.	.
<i>Seseli annuum</i>	.	.	.	.	1	.	1	.	.	.	.	3	4
<i>Seseli campestre</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Seseli dichotomum</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Seseli eriocephalum</i>	3	9	.	.	.	.	.	.	.	.	.	.	.
<i>Seseli gummiiferum</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Seseli hippomarathrum</i>	.	.	.	.	.	9	1	2	.	1	14	.	.
<i>Seseli lehmannii</i>	.	.	.	.	.	.	7	.	.	.	.	.	.
<i>Seseli libanotis</i>	.	1	1	.	1	.	8	3	.	.	9	20	.
<i>Seseli pallasii</i>	.	.	.	.	.	.	.	6	11	.	2	4	4
<i>Seseli rigidum</i>	.	.	.	.	1	.	10	3	.	.	.	.	.
<i>Seseli</i> sp.	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Seseli tortuosum</i>	.	.	1	.	.	.	.	.	.	.	.	.	.
<i>Sesleria heuflerana</i>	.	.	.	.	.	.	13	2	8	1	9	.	.
<i>Setaria pumila</i>	.	.	1	.	1	.	.	.	.	.	.	.	.
<i>Setaria viridis</i>	.	.	.	.	.	.	.	.	3	.	.	.	.
<i>Sherardia arvensis</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Sideritis montana</i>	.	.	.	1	1	8	.	.	.	18	5	.	.
<i>Silene bupleuroides</i>	.	.	.	1	.	1	1	1	.	1	3	.	.
<i>Silene chlorantha</i>	.	.	.	1	1	.	.	.	.	.	.	.	.
<i>Silene cretacea</i>	.	.	7	1	4	.	.	.	.	.	.	.	.
<i>Silene csereii</i>	.	.	.	.	.	.	.	.	.	.	1	.	.
<i>Silene flaves cens</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Silene italica</i>	.	.	.	.	.	.	8	.	.	2	.	2	.
<i>Silene latifolia</i>	.	.	.	.	.	.	1	.	.	1	.	.	.
<i>Silene procumbens</i>	.	.	1	.	.	.	.	.	.	.	.	.	.
<i>Silene</i> sp.	.	.	.	.	.	.	.	.	.	1	5	.	.
<i>Silene viridiflora</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Silene viscaria</i>	.	.	.	.	.	.	.	.	.	.	.	2	.
<i>Silene vulgaris</i>	.	.	.	.	.	.	3	2	.	4	3	7	.
<i>Silene zavadz-kii</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Sisymbrium loeselii</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Sisymbrium polymorphum</i>	3	.	1	.	.	2	.	.	.	.	.	.	.
<i>Solanum nigrum</i>	.	.	.	.	.	.	.	.	2	.	.	.	.
<i>Solidago virgaurea</i>	.	.	.	.	.	.	1	1	.	.	.	.	.
<i>Sonchus asper</i>	.	.	.	1	.	.	.	.	.	.	.	.	.
<i>Sorbus aria</i>	.	.	.	.	.	.	2	3	3	.	.	.	.
<i>Sorbus aucuparia</i>	.	.	.	.	.	.	2	1	.	.	.	.	.
<i>Sorbus dactica</i>	.	.	.	.	.	.	4	.	.	.	.	.	.
<i>Sorbus graeca</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Spiraea chamaedryfolia</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Spiraea crenata</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Spiraea hypericifolia</i>	6	.	.	.	.	.	.	.	.	.	.	.	.
<i>Squamarina cartilaginea</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Squamarina lentigera</i>	.	.	.	.	.	.	.	.	.	.	6	.	.
<i>Stachys germanica</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Stachys officinalis</i>	.	.	.	.	.	.	.	3	.	2	.	.	.
<i>Stipa tirs a</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Stipa zalesskii</i> aggr.	3	.	1	1	1	5	2	.	.	.	1	.	.
<i>Streblotrichum convolutum</i>	.	.	.	.	.	.	.	.	.	.	.	2	.
<i>Syntrichia calcicola</i>	.	.	.	.	.	.	.	.	.	.	1	.	.
<i>Syntrichia montana</i>	.	.	.	.	.	.	.	.	.	.	.	1	.
<i>Syntrichia ruralis</i>	.	.	1	.	3	.	1	.	.	.	.	11	.
<i>Syntrichia ruralis</i> var. <i>ruraliformis</i>	.	.	.	.	.	.	.	.	.	.	1	.	.
<i>Syringa vulgaris</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Tanacetum achillefolium</i>	.	1	.	.	.	.	.	.	.	.	.	.	.
<i>Tanacetum corymbosum</i>	.	.	.	.	1	1	2	3	.	.	.	.	.
<i>Tanacetum kittaryanum</i>	.	3	.	.	.	.	.	.	.	.	.	.	.
<i>Tanacetum millefolium</i>	.	.	.	.	.	2	.	.	.	.	.	.	.
<i>Tanacetum santolina</i>	.	9	.	.	.	.	.	.	.	.	.	.	.
<i>Taraxacum</i> sect. <i>Dioszegia</i>	.	.	1	4	3	8	.	2	.	1	.	.	.
<i>Taraxacum</i> sect. <i>Erythrocarpa</i>	.	.	.	.	.	.	9	.	.	.	.	.	.
<i>Taraxacum</i> sect. <i>Erythrosperma</i>	.	1	.	1	1	.	2	.	.	3	1	11	.
<i>Taraxacum</i> sect. <i>Piesis</i>	6	.	.	1	.	.	.	.	.	.	.	.	.
<i>Taraxacum</i> sect. <i>Scariosa</i>	.	.	1	.	1	.	.	.	.	.	.	.	.
<i>Taraxacum</i> sect. <i>Taraxacum</i>	.	.	.	.	1	3	1	.	.	.	.	9	.
<i>Taraxacum</i> sp.	.	.	.	.	.	.	.	2	.	1	.	2	.
<i>Taxus baccata</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Teesdalia nudicaulis</i>	.	.	.	.	.	.	.	.	.	.	.	2	.
<i>Tephroseris integrifolia</i>	.	.	.	.	.	.	1	6	.	.	.	.	.
<i>Thalictrum foetidum</i>	.	.	.	.	.	.	8	1	3	.	.	.	.
<i>Thalictrum simplex</i>	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Thaloidima sedifolium</i>	.	.	.	.	.	.	.	.	.	.	.	5	.
<i>Thesium alpinum</i>	.	.	.	.	.	.	.	6	.	.	.	2	.
<i>Thesium brachyphyllum</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Thesium linophyllum</i>	.	.	1	.	5	7	17	.	2	1	2	.	.
<i>Thesium procumbens</i>	.	1	.	.	7	.	.	.	.	.	.	.	.
<i>Thesium ramosum</i>	.	3	13	12	16	3	.	.	.	.	.	.	.
<i>Thesium</i> sp.	.	.	.	.	1	.	.	.	.	.	.	.	.
<i>Thlaspi arvense</i>	.	.	1	.	.	.	.	.	.	.	.	.	.
<i>Thymelaea passerina</i>	.	.	.	.	3	.	.	.	.	.	.	.	.
<i>Thymus guberlinensis</i>	.	3	.	.	.	.	.	.	.	.	.	.	.
<i>Thymus moldavicus</i>	.	.	.	.	.	.	.	2	.	.	4	.	.
<i>Thymus</i> sp.	.	.	.	.	.	.	8	.	5	.	.	.	.
<i>Tilia cordata</i>	.	.	.	.	.	.	2	1	1	.	.	.	.
<i>Tilia platyphyllos</i>	.	.	.	.	.	.	1	.	.	.	.	.	.
<i>Tofieldia calyculata</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Toninia coeruleonigricans</i>	.	.	.	.	.	.	.	.	.	.	6	.	.
<i>Toninia</i> sp.	.	.	.	.	.	.	.	.	8	.	.	.	.
<i>Torilis japonica</i>	.	.	.	.	.	.	.	.	.	.	.	2	.
<i>Tortella inclinata</i>	.	.	.	.	.	.	1	.	3	8	2	.	.
<i>Tortula caucasica</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Tortula muralis</i>	.	.	.	.	.	1	.	.	.	1	1	4	.
<i>Tortula</i> sp.	.	.	.	1	.	.	.	.	.	2	.	.	.
<i>Tragopogon dasyrhynchus</i>	.	.	.	.	.	1	.	.	.	.	.	.	.
<i>Tragopogon dubius</i>	.	1	1	.	1	.	.	1	2	5	.	.	.
<i>Tragopogon marginifolius</i>	.	1	.	.	.	.	.	.	.	.	.	.	.
<i>Tragopogon pratensis</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Trichostomum crispulum</i>	.	.	.	.	1	.	.	.	.	.	1	.	.
<i>Trifolium alpestre</i>	.	.	.	.	.	2	2	4	7	2	1	4	.
<i>Trifolium ambiguum</i>	.	.	.	.	.	2	.	.	.	.	.	.	.
<i>Trifolium arvense</i>	.	.	.	.	.	.	1	.	11	10	.	.	.
<i>Trifolium aureum</i>	.	.	.	.	.	.	.	.	.	7	.	.	.
<i>Trifolium campestre</i>	.	.	.	.	.	.	1	1	5	5	.	.	.
<i>Trifolium dubium</i>	.	.	.	.	.	.	.	.	.	1	.	.	.
<i>Trifolium montanum</i>	.	.	.	.	1	.	3	1	2	.	.	.	.
<i>Trifolium pratense</i>	.	.	.	.	.	.	.	1	.	.	.	.	.
<i>Trifolium repens</i>	.	.	.	.	.	.	.	.	.	.	.	2	.
<i>Trifolium rubens</i>	.	.	.	.									